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Isolation and diagnosis of Streptococcus mutans and Streptococcus sorbinus from patients with tooth decay and gingivitis Wassan Lowrance Hassan¹, Abid Ahmad Erdeni², Taghreed Khudhur Mohammed³

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ABSTRACT The Streptococcus bacteria Streptococcus sobrinus and *Streptococcus mutans* are one of the most common types in the mouth, where tooth decay, necrosis and gingivitis are among the health problems that the Iraqi society suffers from, as the treatment of bacteria has become a challenge due to their ability to resist many antibiotics. The study aimed to isolate and diagnose Streptococcus mutans and Streptococcus sobrinus and study their resistance to a number of antibiotics, and evaluate the susceptibility of Streptococcus mutans and Streptococcus sobrinus isolates in biofilms. Using biochemical tests and phenotypic diagnostics. The study included the collection of (275) oral swabs from patients with tooth decay from the area of caries on the surface of the tooth and from cysts for infected patients and visitors at the Medical City Hospital in Baghdad Governorate with ages ranging between (6-70) years and of both sexes for the period from 1/9/2022 to 1/11/2022. the number of isolates was 6 (2.18%) belonging to *Streptococcus mutans* and only one isolate (0.36%) of *Streptococcus sobrinus*, the sensitivity of all the isolates of the bacteria under study was tested towards 10 antibiotics, and the results showed that the isolates were resistant to most of the antibiotics used in this study, and all the isolates were chosen to test their ability to form biofilms using the microtitration plate, and the results showed that most of the isolates were strong in the formation of biofilms.

عزل وتشخيص بكتريا Streptococcus mutans و Streptococcus sorbinus المعزولة من

مرضى تسوس الاسنان والتهاب اللثة

وسن لورنس حسن , أ.م. د عبد أحمد أردينى , أ.د تغريد خضر محمد

الملخص

تعد بكتريا Streptococcus mutans و Streptococcus sobrinus من أكثر الأنواع شيوعا في الفم، إذ يعتبر تسوس الأسنان وتنخرها والتهاب اللثة من المشاكل الصحية التي يعاني منها المجتمع العراقي، إذ أصبح علاج البكتريا تحدياً بسبب قدرتها على مقاومة العديد من المضادات الحيوية . الهدف من الدراسة هو عزل وتشخيص البكتريا وأختبار مقاومتها للمضادات الحيوية و قدرتها على تكوين الأغشية الحيوية بأستخدام الخبارات بايوكميايئية وتشخيصات مظهرية، إذ إن عدد العينات الكلي كان ٢٧ التي عينة تم جمعها من مرضى تسوس وتنخر الاسنان والتهاب اللثة من مراجعين في مستشخيصات مظهرية، إذ إن عدد العينات الكلي كان ٢٧ التي عينة تم جمعها من مرضى تسوس وتنخر الاسنان والتهاب اللثة من مراجعين في مستشخوس منه منه و عزل وتشخيص البكتريا وأختبار مقاومتها للمضادات الحيوية و قدرتها على تكوين الأغشية الحيوية بأستخدام الخبارات بايوكميايئية وتشخيصات مظهرية، إذ إن عدد العينات الكلي كان ٢٧ التي عينة تم جمعها من مرضى تسوس وتنخر الاسنان والتهاب اللثة من مراجعين في مستشفى مدينة الطب في محافظة بغداد وبأعمار تراوحت بين (٢-٧٠) سنة ومن كلا الجنسين للمدة من ١٩/١٩ ٢٠٢ الى من مراجعين في مستشفى مستشفى مدينة الطب في محافظة بغداد وبأعمار تراوحت بين (٢-٧٠) سنة ومن كلا الجنسين للمدة من ١٩/١٩ ٢٠ الى الماد التي عينة تم جمعها من مرضى تسوس وتنخر الاسنان والتهاب اللثة من مراجعين في مستشفى مدينة الطب في محافظة بغداد وبأعمار تراوحت بين (٢-٧٠) سنة ومن كلا الجنسين للمدة من ١٩/١٩ ٢٠ المار المالي التها التي عينة تم جمعها من مرضى تسوس وعلي أول ترامي من مراجعين في من مراجعين في مستشفى مدينة الطب في محافظة بغداد وبأعمار تراوحت بين (٢-٢٠) سنة ومن كلا الجنسين للمدة من ١٩٧٦ الى الالمالي الماليان والتها أول من مراجعين في ماليات أن عدد العزلات ٦ (٢٠,١٧٪) التي تنتمي إلى معلمه من السنان وعزلن المالية عنها مرابي وي الدراسية على ما محسولة واحدة فقط (٣٠,٠٠٪) تعود الى الاراليان ولي المارين الماليان والتها وي بالتفريا علي العار اليان والتها وي الماليان والته من الماليان ولالياليا ولالي ف بكتريا معادم معلم المضادات العزلات ٦ (٢٠,١٧٪) التي تنتمي ولى معظم العزلات كانت قوية في على معلميا على تكوين الأعشية العريات العلي وي العلي وي العن ماليا ومن ولال كانت قولة معلم المارم ولالياليا مالياليا ماليان والياليان والنان واللام

Introduction: Pathological periodontal disease is one of the most prevalent diseases in the world: dental caries, tooth decay and periodontal disease, and gingivitis [1]. Dentistry leads to the disintegration of the enamel layer or acid-mediated root surface damage, mainly due to diet, causing dental causes and other oral diseases [2] and damage to the supporting tissues of the teeth [3]. *Streptococcus mutans* is one of the most important causes of dental marketing. It is a Gram-positive, facultative anaerobe. This high profile is shown to influence biofilm formation, acid production, and salt and acid tolerance [4,5]. Citrus fruits also contain carbohydrates such as glucose and sucrose. It also contributes proteins to strep bacteria. mutans such as glycosyltransferases-GTFs, biomembrane regulatory proteins, wall-associated protein-coupling proteins, and glucan-experiment proteins, cause mutations in *Streptococcus* spp. and form caries-lesions [6,7]. In association with *Streptococcus sobrinus*, which is considered one of the types of bacteria that cause diseases of rapture, it is considered an anaerobic bacterium, spherical in shape, gram-positive, and its optimal growth temperature is at 37 C° in environments with low pH 6.3. Human oral is home to *Streptococcus* [8]. The aims of the study are to study the diagnosis and identification of *Streptococcus mutans* and *Streptococcus sobrinus* and to study their resistance to antibiotics. Also, the formation of biofilms.

Materials and methods

- Sample collection: The study included the collection of (275) oral swabs from patients with tooth decay from the area of caries on the surface of the tooth and from cysts for infected patients and visitors at the Medical City Hospital in Baghdad Governorate with ages ranging between (6-70) years and of both sexes for the period from 1/9/2022 to 1/11/2022. Transport Medium is a brain-heart infusion broth and the samples were then transferred directly to the laboratory for the implantation of swabs on the rich and selective culture media, and then incubated in the incubator for a period of 24 hours at a temperature of 37 C° for diagnostic tests later.
- Culture media:
 - 1. Enrichment media) Blood agar): was used to test the ability of bacteria to lyse blood and to know the type of haemolysis.
 - 2. Simple media (Nutrient agar medium): was used to test the ability of bacteria to grow on it.
- 3. Urea agar was used to test the ability of bacteria to produce the enzyme urease.
- 4. Motility test medium was used to study the possibility of bacteria on movement.
- 5. Mitis Salivarius Bacitracin AGAR (MSBA): It is the selective medium of *Streptococcus mutans* prepared according to [9,10].
- 6. Voges-Proskauer and methyl red and Indole and Simmmons Citrate: This medium was used to detect the complete decomposition of sugars and the production of organic acids after 24 hours (MR test) or partial decomposition of sugars and the production of acetone after embrace for 48 hours (VP Test) and the results of Indol and simmonus Citrate test were negative [11].
- 7. Brain-Heart infusion broth: It has been used as a medium to ensure that bacteria are maintained until they reach the laboratory [11].
- Collection and Cultivation of Sample: Samples are transplanted directly on Blood agar, and Mitis Salivarius Bacitracin agar (MSBA) and were embraced under intuitive conditions at 37 °C for 18-24 hours, and then saved at 4 C° until diagnostic tests were conducted, the diagnosis was conducted according to the standard methods used for that and contained in [12].

- **Bacterial Identification:** To distinguish between colonies of *Streptococcus mutans* and *Streptococcus sobrunis*, MSBA medium was used. Then, 10% Mannitol and 4% Triphenyl tetrazolium chloride (TTC) were added by diffusion method. and that the color of the medium changed to dark pink, which is evidence of the reduction of (TTC) and the decomposition of mannitol sugar by Mannitol-1-phosphate dehydrogenase [13,14,15].
- **Biochemical tests:** The tests (Oxidase Test, Catalase Test, IMViC Test, Urease Test, Motily Test, Coagulase Production enzyme, and Lactose Fermentation) were conducted according to the way that each of [16,17].
- Microscopic examination by using Gram stain.
 - Identification by VITEK 2 [18] .
- Antibiotics Susceptibility Test According to CLSI: A bacterial sensitivity test for ten antibiotics (Bioanalyse, Turkey) mentioned in Table (1) was performed where all bacterial insolation is studied against antibiotics by Disc Diffusion Method using the Kirby Bauer method [19,20].

μg/disk	code	Antibiotics	
15	AK	Amikacin	-1
15	AZM	Azithromycin	-2
30	CFM	Cefixime	-3
30	CTZ	Ceftazidime	-4
30	CRO	Ceftriaxone	-5
30	CAZ	Cephalexin	-6
5	CIP	Ciprofloxacin	-7
5	IMI	Imipenem	-8
5	LEV	Levofloxacin	-9
15	TBO	Tobramycin	-10

Table 1: Antibiotics and their concentrations

• **Detection of biofilm formation:** Microtiter Plate (MTP) was used The optical density readings (OD) were scheduled using ELISA with a length of 630 nm [21].

Results

The results showed that 7 isolation and (2.54%) belong to *S. mutans* and *S. sobrinus*, as it was found that 6 isolation (2.18%) dates back to *S. mutans*, while only one isolation (0.36%) was isolated due to *S. sobrinus* (Table 2).

(%) Total	(%) negative samples	positive samples on (%) bacteria	Type of bacteria
(100) 275	(97.45) 268	(2.18) 6	S. mutans
		(0.36) 1	S. sobrinus
		(2.54) 7	(%) Total

Table (2) percentages of bacterial isolates isolated from patients

Current results showed that the percentage of bacteria is higher in females than in males, with 4 (1.45 %) females with *S. mutans*, 1 (0.36 %) female with *S. sobrinus* (Table 3). In the males, 2 (0.72 %) of *S. mutans*, and did not isolate *S. sobrinus* in males. It was also the highest rate of *S. mutans* within the age group 0-10 years, followed by the age group 11-20 years and 61-70 years, with an insulation rate of 3 (50%), 2 (33.33%) and 1 (16.66%), respectively, table (4). As for *S. sobrinus* has been isolated from an 8 -year -old girl, who was suffering from tooth decay with dental deformation and gums.

 Table (3) Percentage of bacterial isolates in both males and females suffering from caries and necrosis

 Teeth and gingivitis

males and females (%)	Total (%)	Negative samples in females (%)	Positive samples in females (%)	Total (%)	Negative samples in (%)males	positive samples in (%) males	Type of bacteria
440	۱۷.	١٦٦	(1,£0) £	1.0	۱۰۳	۲ (۲۷,۰)	Strep.
(\cdots)	(٦١,٨١)	(٦٠,٣٦)		("^,1^)	(۳۷, ٤٥)		mutans
7 7 0	18.	(71,50) 179	۱ (۳۳, ۰)	1.0	1.0	(·) ·	Strep.
$(\cdot \cdot \cdot)$	(11,41)			(٣٨,١٨)	(٣٨,١٨)		sobrinus

Table (4) Distribution of Streptococcus mutans bacteria by age groups in patients suffering from	n dental
caries, necrosis and periodontitis	

(%) Total	Isolates of Strep. mutans in females (%)	Isolates of Strep. mutans in males (%)	Age group
۳ (۰۰)	(0,) 1	(••) /	10-0
(٣٣,٣٣) ٢	((0))	(••) 1	20-10
(•) •	(•) •	(·) ·	30-21
(•) •	(·) ·	(·) ·	40-31
(•) •	(•) •	(·) ·	50-41
(•) •	(•) •	(·) ·	60-51
(17,77) 1	(* 0) 1	(·) ·	70-61
۲ (۱۰۰)	(1) 2	(1) 4	(%) Total

The results of the study were somewhat compatible with local and international studies. The researcher Flayyih *et al Streptococcus mutans* isolated from primary school students and from patients with private clinics auditors in Baghdad was 15 of the totals of 109 patients with a percentage of percentage (13.79%) using the different selective and rich media to isolate *Streptococcus mutans*. The highest insulation rate for females was compared to males within different age groups, (49.3%) in the age group of females (4-12) years, and (39.3%) in the age group of males (4-12) years [22].

The Identification was based on the form of bacterial isolates that were developed on Enriched, Special and Selective cultures, *S.mutaus* colonies appeared on the Nutrient agar medium in the heart and brain agar extract (BHIA) Heart and brain agar extract in a white color tending to light grey with diameters ranging between 2-1 mm, while *S. sobrinus* colonies also appeared white tending to light gray but slightly larger than *S. mutans* most of their diameters ranged between 5-2 mm (Figures 1 and 2), and the growth of both types of bacteria under anaerobic conditions (5%CO₂) was much better when incubating bacteria under aerobic conditions. This matches previous research findings[22,23,24]. On the medium of blood agar, the two species gave *S. mutans* and *S. sobrinus* is the hematolysis of the Gamma hemolysis type γ , and this is evidence of the inability of bacteria to produce haemolysin, Figure (3), and these results were consistent with what was stated in previous studies [25].



Figure 1: Bacteria colonies S. mutans on nutrient agar medium



Figure 2: colonies of S. sobrinus on Nutrient agar medium



Figure (3) A- colonies of *S*.*mutans* on blood agar media after 48 h incubation, γ-hemolysis. B- colonies of *S*. *sobrinus* on blood agar media after 48 h incubation, gamma-γ-hemolysis.

For the growth of *Streptococcus mutans* and *Streptococcus sobrinus* on MSBA, bacterial colonies appeared dark blue to pink with light-colored edges after spraying the medium with a 4% solution of triphenyl tetrazolium chloride and aerobic incubation at 37°C for an additional hour. Only the *S. mutans* colonies. are smaller than *S. sobrinus* colonies, Figures 4 and 5.



Figure(4) colonies S. mutans and S. sobrinus on MSBA medium prior to spraying of Tetrazolium chloride (aerobic incubation at 37°C, for an additional hour)

Tikrit Journal of Pure Science (2024) 29 (1): 37-48 Doi: <u>https://doi.org/10.25130/tjps.v29i1.1471</u>



Figure 5: colonies S. mutant and S. sobrinus on MSBA medium

Microscopy using the Gram stain shows that all bacterial isolates were in the form of cocci tending to the oval positive for the Cram stain, resulting in the form of short chains or pairs (Figure 6). These results are consistent with previous local and international studies [26,27].



Figure (6) *Streptococcus mutans* positive bacteria under a normal light microscope using an oil lens at full magnification (X1000).

Diagnosis of isolates using biochemical Tests: As for the IMVIC test, which included the Indole test, the Methyl red (MR) test, the Vogus-Proskauer (VR) test and the consumption of citrate, the results for the *S. mutans* and *S. sobrinus* isolates showed that they were negative for the Indole, MR and citrate consumption. While a positive result for the VR test gave an indication that the bacteria produced acetoin after the fermentation of glucose sugar Table (5) and Figure (7). After biochemical tests, it was found that all *S. mutans* and *S. sobrinus* isolates had given a negative result for the Catalase test Figure(8). The isolates also showed a negative result for the Oxidase test, [28,29].



Figure (8) negative result of Isolate No. 2 of S. mutans and S. sobrinus in the catalase test

As for the production of bacterial isolates of the enzyme urease, the results showed that they were unable to produce the enzyme urease, so gave a negative result for this test. To confirm the diagnosis of isolates as *S. mutans and Sterp. sobrinus*, they were also examined by the VITEK2 system (Table 6) The results showed that the isolates *of Strepr. mutans* gave positive results for the test of TyrA, LAC, dSOR, dMAN, SAL, AGAL, dMNE, SAC, dMAL, dGAL, Dtre BACl. AGLU, LeuA, AlaA, NOVO, dRAF, OPTO, for *S. sobrinus*. The same results as *S. mutans* isolates except that dSOR had a negative result (Table 6). The results of Abo Baker *et al.* showed that *S. mutans* isolated from patients in the Arab Republic of Egypt with tooth decay and decay had the ability to ferment four main sugars used in their diagnosis: mannitol, sucrose, sorbitol and inulin [30], as well as the two types of bacteria *S. mutans* and *S. sobrinus* have the ability to ferment and consume lactose, mannose

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and raffinose, and the production of dextran of sucrose has an effective role in the pathogenesis of bacteria and the cause of decay. While it does not have the ability to consume [31,32].



Figure (7) IMVIC biochemical tests (Negative result of A: indole, B: methyl red, D: citrate consumption), C: positive result of Voges-Proskauer test

Table (6) Phenotypic and biochemical tests for S. sobrinus and S. mutans

S.sobrinus	S.mutans	Tests	
Spherical, Gram-positive	Spherical, Gram-positive	Gram stain	-1
cocci	cocci		
-	-	Catalase test	۲_
-	-	Oxidase test	-3
-	-	Indole	-4
-	-	Methyl Red	_0
+	+	Voges-Proskauer	۲_
-	-	citrate consumption	-7
-	-	Urease	_A
+	+	(LeuA) Leucine arylamidase	-9
+	+	(AGLU) Alfa-glucosidase	-10
+	+	(ALaA)	-11
+	+	(NOVO) Novobiocin Resistance	-12
+	+	(dRAF) D-Raffinose	-13
+	+	(OPTO) Optochin Resistance	-14
+	+	(TyrA) Tyrosine arylamialase	-15
	+	(dSOR) D-Sorbitol	-16
+	+	(LAC) Lactose	-17
+	+	(dMAN) D-Mannitol	-18
+	+	(SAL) Salicin	-19
+	+	(AGAL) Alpha-galactosidase	-20
+	+	(dMNE) D-Mannose	-21
+	+	(SAC) Saccharose /Sucrose	-22
+	+	(dMAL) D-Maltos	-23
+	+	(dTRE) D-Treahlose	-24
+	+	(dGAL) D-Galactose	-25
+	+	(BACL) Bacitracin Resistance	-26
•	-	Motility	_ T V
-	-	Coagulase	-28

susceptibility of S. mutans and S. sobrinus to antibiotics

The bacterial sensitivity test for antibiotics was performed using Muller-Hinton Agar medium and ten types of antibiotics. The current study showed that bacterial isolates show a difference in their resistance and sensitivity to the antibiotics under study. Fig. 10 and Appendix 3 showed that the *S. sobrinus* isolate was moderately sensitive



to Imipenem and sensitive to Amikacin, Tobramycin, Ciprofloxacin, Azithromycin, and Levofloxacin, while it was resistant to Ceftazidime, Cephaloxin, Cefixime and Ciprofloxacin. With regard to Strep.mutans, the results showed that (6) isolates have high resistance to Ceftazidime, Cefixime, Ceftriaxone and Cephaloxin by (100%), followed by Tobromycin 5 (83.33%), Figure (9) Table (7). While bacterial isolates recorded the highest percentage of sensitivity to Amikacin and Ciprofloxacin (100%), followed by Imipenem (83.33%), then Azithromycin and Levofloxacin (66.66%) respectively.



Figure (9) Resistance sensitivity of some isolates of *S. mutans* and *S. sobrinus* to a number of antibiotics (On MHA at 37 for 24 hrs)

Sensitive isolates (S)		Intermediate isolates (I)		Resistant isolates (R)		Cod	Antibiotics
Percentage %	Number	Percentage %	Number	percentage %	Number		
1	٦	•	•	0	0	AK	Amikacin
11,11	٤	17,77	١	16.66	1	AZM	Azithromycin
0	0	•	•	100	6	CFM	Cefixime
0	0	•	•	100	6	CAZ	Ceftazidime
0	0	•	•	100	6	CRO	Ceftriaxone
100	6	•	•	0	0	CIP	Ciprofloxacin
83.33	5	13,33	١	0	0	IMI	Imipenem
66.66	4	17,77	1	16.66	1	LEV	Levofloxacin
0	0	•	•	100	6	CN	Cephalexin
16.66	1	•	•	83.33	5	TOB	Tobramycin

Table (7) Susceptibility of (6) isolates of S. mutans to antibiotics

R: Resistance, I: Intermediate, S: Sensitive

It was observed that the sensitivity of the isolates S. mutans and S. sobrinus is sensitive to some antibiotics under study, and these results are close to the results of some other studies carried out by a number of researchers. The results of the current study are consistent with previous studies, including the local results reached by Saleh with other researchers in 2023, where they showed isolates of S. mutans isolated from Baghdad patients were sensitive to Ciprofloxacin 100%). It was resistant to the anti-Cefixime by (100%) [33] and these results were identical to the results of the current study, and the results reached by the researchers Saleh and Abdel-Rahman in 2017 were consistent with the current study, where it appeared that the isolated S. mutans isolates from patients in Thi Qar Governorate were sensitive to anti-Imipenem by (78.3%) and Ciprofloxacin by (60.9%) and the study did not agree with the Ceftriaxone because its results were sensitive to bacteria (60.9%) While it was resistant by (100%) in the current study, the reason may be due to the size of the sample and the number of positive samples, as well as for the antibody Amikacin, which showed resistance, while in the current study, the bacteria were sensitive to it by (100%), the reason may be due to the indiscriminate use of antibiotics [34] The current study agreed with the results of a study in Argentina carried out in 2021 by researcher Bachmeier et al., that the bacterial isolates of S. mutans were sensitive to Amikacin (83.30%) and Ciprofloxacin (77.75%) [35]. As for S. sobrinus, the results showed that that isolation, which was only one isolation that was isolated during the study, had resistance to the antibodies Ceftazidime, Cephalexin, Cefixime and Ceftriaxone, while Amikacin, Tobramycin, Azithromycin and Levofloxacin were

TJPS

sensitive and were moderately sensitive to Imipenem, and because of the rarity of their isolation, they are present in small proportions in patients with caries and tooth decay, so studies on them are rather few, and the researcher Salman pointed out in 2015 in a study conducted on the sensitivity of *S. sobrinus* as being sensitive to ampicillin and Penicillin antagonists and less sensitive to chloramphenicol and Cefazolin [36] One of the main reasons for bacterial resistance to antibiotics is the horizontal transfer of genes, as it is the reason for the development of the ability of bacteria to resist pesticides and antibiotics through the mechanism of gene transfer from bacteria to other bacteria through plasmids or through viruses known as phages or through sexual filaments [37].

Susceptibility of bacteria to Biofilm formation

In Figure (10) it was noted that 4 isolates (1, 3, 4, 5) have a high ability to form biofilms, while isolates (2, 6) have a moderate ability to form membranes, as for the isolation of S. sorbinus showed the ability to form biofilms strongly, Table (9) showed that the total number of isolates 7 (100%) of bacterial isolates showed the ability to form membranes at varying rates, the results of the examination of the microplate plate of researchers Zayed et al. in 2021) showed that (80) isolates were isolated from S. mutans from dental plaque samples have the ability to form higher amounts of strong biofilm patterns compared to saliva samples, evidenced by the strong production of biofilm by 35 isolated isolates of dental plaque compared to only 26 isolated from saliva. Also, 4 (5%) isolated isolates of dental plaque showed an ability to produce moderate biofilm versus 12 (15%) isolates isolated from saliva samples. and one isolation (1.25%) isolated from dental plaque showed impairment in the ability to produce biofilms compared to two isolates (2.5%) of isolated isolates from the saliva sample. Comparing the data obtained from isolated samples from saliva and dental plaque samples, it was observed that isolated isolates from dental plaque have the ability to form a stronger biofilm than those isolated from saliva samples [38]. The findings of Alhasani et al. in 2020 were somewhat similar to those of the current study, as 261 isolates of bacteria were tested S. mutans to see how capable they are to form biofilms, where the results showed 31 isolates (2). (12%) had the ability to form strong biofilms, 46 isolates (18%) had an average biofilm formation capacity, and 184 isolates (72.2%) had a poor biofilm formation capacity [40].



Figure (10): Isolates of *S. mutans* (5, 4, 3, 1) have a high ability to form biofilms, while isolates (2, 6) have a medium ability to form membranes, while isolates of *S. sorbinus* was shown to have the ability to form biofilms in a strong way. C-: Negative Control, C+: Positive Control.

T = 1 = (0)		10 11	1 41	- C '4 1. '1'4	4 - C	1
1 9 DIE (9)* 1	NUMBER OF N. <i>mutan</i>	s and N sorninus and	i the nercentades	or its anility	to torm	niorume
\mathbf{I} and (\mathcal{I})		, and D. Sviviins and	i the percentages	or no aome	to to m	DIOLIHIS

Percentage(%)	Number of isolates (N)	susceptibility of bacterial isolates Formation of biofilms
۷۱,٤۲	5	Strong
28,08	2	Moderate
0	0	Weak
0	0	No Biofilm
100	7	Total



Conclusion

The results of the current study showed that the percentage of isolation of S. mutans and S.sobrinus isolated from the mouth 2.54%. and Isolates are characterized by their antibiotic constituent (Cefixime, Ceftazidime, Ceftriaxone, Cephaloxin, Tobramycin). and less resistant (Levofloxacin and Azithromycin). And The current study showed that all isolates had the ability to form biofilms differently, where 71.42% had the ability to form strong biofilms and 28.57% formed medium-strength biofilms.

References:

[1] Garcke, H. and Nürnberg, R. (2021). Structure-preserving discretizations of gradient flow for axisymmetric two-phase biomembranes. IMA *Journal of Numerical Analysis*, 41(3): 1899-1940.

[2] Elgamily, H. M.; Gamal, A. A.; Saleh, S. A.; Wahab, W. A. A.; Hashem, A. M.; and Esawy, M. A. (2019). Microbiological and environmental assessment of human oral dental plaque isolates. *Microbial pathogenesis*, 135: 103626.

 [3] Poza-Pascual, A., Serna-Muñoz, C., Pérez-Silva, A., Martínez-Beneyto, Y., Cabello, I., & Ortiz-Ruiz, A.
 J. (2021). Effects of Fluoride and Calcium Phosphate-Based Varnishes in Children at High Risk of Tooth Decay: A Randomized Clinical Trial. International journal of environmental research and public health, 18(19): 10049.

[4] Taketo. K.; Naoki, N.; Saori, Y.; Yoshiaki, T.; Jun. I.; Yasutaka. H.; Hidenobu. S.(2016). Inhibition of *Streptococcus mutans* biofilm formation.

[5] Lemos, J. A.; Palmer, S. R.; Zeng, L.; Wen, Z. T.; Kajfasz, J. K.; Freires, I. A.; Abranches, J. and Brady, L. J. (2019). The Biology of *Streptococcus mutans*. Microbiology spectrum, 7(1), 10.1128/microbiolspec.GPP3-0051-2018.

[6] **Matsumoto-Nakano**. (2018). Role of *Streptococcus mutans* surface proteins for biofilm formation Japanese Dental Science Review .

[7] Salehi,B.; Kregiel,D.; Mahady.G.; Sharifi-Rad.J.; Martins.N.; Rodrigues.C . (2020). Management of *Streptococcus mutans-Candida spp*. Oral Biofilms' Infections: Paving the Way for Effective Clinical Intervention. *J Clin* . 9(2): 517.

[8] Sales, MJ.; Herbert, WG.; Du, Y.; Sandur, AS.; Stanley, NM.; Jensen, PA. (2018). Complete genome sequences of *Streptococcus sobrinus* SL1 (ATCC 33478 = DSM 20742), NIDR 6715-7 (ATCC 27351), NIDR 6715-15 (ATCC 27352), and NCTC 10919 (ATCC 33402). Microbiol Resour Announc 7:e00804-18.

[9] Abd, Suha T., and Abbas F. Ali. (2016) "The Effect of Zinc Oxide Nanoparticles on Streptococcus mutans of Human Saliva (In Vitro Study)." *Journal of baghdad college of dentistry* 28.2: 158-164.

[10] **Nagamine, Y., Hasibul, K., Ogawa, T., Tada, A., Kamitori, K., Hossain, A., and Miyake, M.** (2020). D-Tagatose Effectively Reduces the Number of Streptococcus mutans and Oral Bacteria in Healthy Adult Subjects: A Chewing Gum Pilot Study and Randomized Clinical Trial. Acta Medica Okayama, 74(4), 307-317.

[11] **MacFaddin, J.F.** (2000). Biochemical test for identification of medical bacteria .Lippincott Willims and Wilkins. Philadelphia, USA.

[12] Forbes, B.A. ; Sahm, D.F. ; and Weissfeld, A.S. (2007). Bailey and Scott's diagnostic microbiology. 12th ed, evolve, china.

[13] Al-Mudallal, N. H., Al-Jumaily ,E. F., Muhimen. N. and Al-Shaibany A. A.(2008). Isolation and Identification of Mutans Streptococci Bacteria from Human dental Plaque samples. Journal of Al-Nahrain University Vol.11(3), pp.98-105.

[14] Levinson, W. (2016). Review of Medical Microbiology and Immunology. 14thed. McGraw-Hill education, Inc. PP 821.

[15] **Wanger, A.; Chavez, V.; Huang, R.; Wahed, A.; Dasgupta, A. and Actor, J.K**. (2017). Microbiology and Molecular Diagnosis in Pathology: A Comprehensive Review for Board Preparation, Certification and Clinical Practice. Elsevier 285.

[16] **Brooks, G.F.; Carroll, K.C; Butel , J.S.; Mores, S.A. and Mietzner, T.A.** (2018). Jawetz , Melnick and Adelbergs Medical Microbiology .25th ed. thr McGraw-Hill companies ,United states of American .

[17] Leber, A.L.(2016). Clinical Microbiology Procedures Handbook, 4th ed, vol 2. Washington DC: ASM Press.

[18] Harvey, R.A.; Cornelissen, C.N. and Fisher, B.D. (2013). Lippincott's Illustrated Review Microbiology. 3^d ed. Lippincott Williams and Wilkins Wolters Kluwer business. USA .

[19] **Bauer, A. T.** (1966). Antibiotic susceptibility testing by a standardized single disc method. *Am J clin pathol*, *45*, 149-158.

[20] **Clinical and Laboratory Standards Institute (CLSI).** (2015). Performance Standards foAntimicrobial Susceptibility Testing; Twenty-Fifth Informational Supplement. Approvedstandard M100-S25. CLSI, Wayne, PA.

[21] **Kirmusaoglu, S.** (2019). The Methods for Detection of Biofilm and Screening Antibiofilm Activity of Agents. In ExopolysaccharidesMethods of Preparation and Application. In technology Open. DOI: http://dx.doi.org / 10.5772 / intechopen.84411

[22] Flayyih, A.S.; Hassani, H.H. and Wali, M.H. (2016). Identification of *Streptococcus mutans* from human dental plaque and Dental caris using 16SrRNA gene, Iraqi J. Of science; 57 (1C): 552-557.

[23] **Mozaal,H and Chlelab,R** .(2019). Isolation and Iodenfication of *Streptococcus mutans* from oral infection and testing their sensitivity to some plant extractions. Maaysan Research Journal, 14 (28):256.

[24] **Pisarska,A.; Wolinowska,R.; Rudnicka,J.; Iwanicka-Grzegorek,A.** (2022). Characteristics of Clinical Isolates of *Streptococcus mutans*. J. *MDPI Applied Sciences*. *12*(9), 4579.

[25] Zeng,Y.; Fadaak,A.; Alomeir,N.; Wu,Y .; Wu,T.; Qing,S.; Xiao,J. (2023). Effect of Probiotic *Lactobacillus plantarum* on *Streptococcus mutans* and *Candida albicans* Clinical Isolates from Children with Early Childhood Caries. Int J Mol Sci. 24(3): 2991.

[26] **Saleh, M.; Abd Al-Zahra, H**. (2019). Isolation and identification of *Streptococcus mutans* from dental caries patients at Thi-Qar province/Iraq. University of Thi-Qar Journal of Science, 6(4), 22–27.

[27] Zadeh, E.T.; Aref, P.; Askarizadeh, N.; Emadi F.(2023) In vitro Antimicrobial Effect of Punica granatum Extract versus Chlorhexidine on Streptococcus sobrinus, Streptococcus sanguinis, and Candida albicans. J Res *Dent Maxillofac* Sci. .8(1):18-27.

[28] Abranches, J.; Zeng, L.; Kajfasz, J, *et al.* (2018). Biology of oral Streptococci, Mi crobiol. Spectr.; 6(5) : 1-18.

[29] **Hussein,Sh**.(2020). Study of the diagnosis and isolation of bacteria associated with dental caries in pregnant women in Baghdad provice, EurAsian J . of Bio Sciences; 14 (1) : 1-7 .

[30] **Abo Bakr, R.; Tawfick, M.; Mostafa, Z.; Abdulal, I** A.(2022). Prevalence and Antibiogram of Streptococcus mutans in Dental Plaque and Caries Samples. Azhar International Journal of Pharmaceutical and Medical Sciences. 2022; 2 (2): 83-93, doi: 10.21608/AIJPMS.2022.87671.1084

[31] Salh,A .; Risan,M . and Jasim,H . (2022). Biochemical characteristics and antibiotics susceptibility of *Streptococcus mutans* isolates from dental caries in baghdad city, International J. of Advanced Biological and Biomedical Research; 10, Issue 1: 32-34.

[32] Maasi,G.; Štšepetova,J.; Jõesaar,M.; Olak,J. and Mändar,R. (2022). Different Patterns of Virulence Genes in *Streptococcus mutans* and *Streptococcus sobrinus* Originating from Estonian Toddlers—Mothers Cohort. *Microbiol. Res.* 13(4), 928:936.

[33] **Saleh,B.H.; Ibrahim,R.N.; Al-Ugaili,D.N**. (2023). The Effect of Diode Laser on Viability and Antibiotic Sensitivity of StreptococcusmutansIsolated From Dental Caries. Iraqi Journal of Science. Vol. 64, No. 2, pp: 583-593DOI: 10.24996/ijs.2023.64.2.8

[34] **Aabdl-Rahimea, A.M and Saleh.N.F** .(2017). Evaluation of some antibiotics resistance of different oral bacteria types isolated from patients with gingivitis and periodontitis. University of Thi-Qar Journal Of Science (UTsci). Volume 6, Number 3.

[35] Bachmeier, E.; Goitea, M.E.; Linares, J.; Wietz, F.M.; Jarchum, S.; Jarchum, G.; Brunotto, M.; Mazzeo, M. (2021). Determination of Streptococcus sp. and Candida albicans in the oral cavity of patients undergoing bone marrow transplantation. Journal of *Oral Research*. Vol(10) N°. 6. págs. 1-10.

[36] **Salman, H.A.; Senthikumar, R.** (2015). Identification and Antibiogram Profile of *Streptococcus mutans and Streptococcus sobrinus* from Dental Caries Subjects. Journal of Applied Pharmaceutical Science. Vol.5 (06), pp. 054-057

[37] **OECD**, Safety Assessment of Transgenic Organisms, Volume 4: OECD Consensus Documents, 2010, pp.171-174.

[38] **Zayed,S,M.; Aboulwafa,M.M..; Hashem,A.M and Saleh,S.S** .(2021). Biofilm formation by Streptococcus mutans and its inhibition by green tea extracts, *AMB Express.*; 11: 73. doi: 10.1186/s13568-021-01232-6.

[40] **Alhasani,A.H.; Ishag,R.A.; Al-Akwa,A.A.; Al Shamahy,H.A.W.; M.A.** (2020). Association between the streptococcus biofilm fromation and dental caries experience and antibiotics resistance in adult females. Universal Journal of Pharmaceutical Research; 5(6):18-^Y^r.