# Isolation and Identification of *Candida spp*. In immunocomparomised patients and Detection some virulence factors.

Milad A. Mezher<sup>1</sup>, Wa'ad M. Ra'oof<sup>2</sup>, Khalil I. Bandar<sup>3</sup>

<sup>1</sup>Department of Biology, College Of Education For Pure Sciences, Tikrit University, Tikrit, Iraq

<sup>2</sup> College Of Pharmacy, Tikrit University, Tikrit, Iraq

<sup>3</sup> Department of Biology, College Of Science, Tikrit University, Tikrit, Iraq

### Abstract

One hundred thirty patients undergoing chemotherapy for different types of cancer attending Tikrit Teaching Hospital from January 2013 to Augusto 2013 were enrolled in this study. Their age were range between "11-83 years old". 68 blood samples and 62 sputum samples were collected in this study.

*Candida spp.* Infection was 14(20.5%) isolates from patients blood and 5(8.1%) isolates from patients sputum, were infections more in blood *C.albicans* 8(57.8%), *C.glabrata* 3(21.4%) *C.krusei* 2(14.3%) and *C.parasillosis* while isolates from sputum were 3(60%) of *C.albicans* and *C.glabrata* 2(40%). The isolated were identified according morphological, Cultural and biochemical characteristic.

Fungal isolates in present study produced various virulence factors hemolysin and protease production were produced in *C.albicans* 8(72.8), 11(100%) respectively, while *C.glabrata* hemolysin production were 4(80%) and (40%) of protease production, while *C.parasillosis* isolate were produced protease and hemolysin. C.krusei were produce only hemolysin 1(50%).

# Introduction

Medical mycology education crisis after 1970s, the incidence increased of fungal diseases in immunocompromised hosts stimulated awareness and growth in medical mycology <sup>(1)</sup>.

More than 200 species that have been described, Candida is ubiquitous. Some species are normal habitants of the human microbiological flora of the skin, as well as the gastrointestinal, genitourinary, and even respiratory tracts <sup>(2,3)</sup> but only 10% are known to cause infections in humans <sup>(4,5)</sup> Five most common species (as Candida albicans, C.glabrata, C.parapsilosis, C.tropicalis and C.krusei) cause 95-97% of human candidemias  $^{(4,6)}$ . The remaining 3–5% of Candida blood stream infections are caused by 12-14 different species, and although these are considered rare causes of candidemias, several have occurred in nosocomial clusters and exhibit innate or acquired resistance to one or more established antifungal agents<sup>(2)</sup>. Candida infections of the latter category are also referred to as candidemia and severelv usually confined to immunocompromised persons , such as cancer, transplant and AIDS patients <sup>(7)</sup>. , this yeast exhibits a number of different morphological forms under different environmental conditions; such forms yeast include budding cells (blastospores, blastoconidia), pseudohyphae (elongated cells which appear as filamentous cell chains), true hyphae, and clamydospores <sup>(9)</sup>. Candida species are possess a number of virulents factors (8) such as phospholipase , hemolysis and protease. That helps fungi in nutrient uptake, tissue invasion, adherence and dissemination <sup>(10)</sup>. Hemolysin in red blood cells has also suggested to provide a survival strategy for fungi during opportunistic infections. In Candida, the secretion of hemolysin coupled with iron uptake facilitates hyphal invasion during disseminated candidiasis Proteases are a group of enzymes, whose catalytic function is to hydrolysed peptide bonds of proteins

and break them <sup>(12)</sup>. Secretion of proteinases by pathogen is mandatory in order to degrade the tissue barriers and obtain nutrition at the infection site <sup>(13)</sup>.

Extracellular proteases may play a role in adherence and survival of the pathogen on mucosal surface  $^{(14)}$ , invasion of host tissues  $^{(15,16)}$  and digestion of immunoglobulins  $^{(16,17)}$ . there are a number of publications investigating protease production in *Candida* spp. isolated from different sources  $^{(18)}$ .

Several researchers have reported that production of Secreted aspartyl proteinases is also correlated with hyphal formation, adherence, and phenotypic switching <sup>(19,20)</sup>.

this study aimed to :

• To isolation and identification of candida species infection from immunocompromised patients. By using conventional methods (BHI agar/broth, colony morphology, germ tube test).

• To screening the ability of candida species isolates to produce , protease, hemolysin as a virulence factors.

# Materials & methods:

# Samples Collection:

Sputum and blood samples were collected from 130 patients undergoing chemotherapy their age varried "11-83 years old", for different types of cancer attending Tikrit Teaching Hospital from January 2013 to Augusto 2013.

Blood samples were cultured on BHI agar/broth for isolation media according Glenn& John (1975)<sup>(21)</sup> and Sputum samples were cultured on (SDA).

1. Identification Of Candida Isolates:

- Germ Tube Test was determined according to Binesh & Kalyani  $(2011)^{(22)}$ .

- Chlamydospore Formation all *Candida* isolates were tested for the production of chlamydospores in corn meal agar (CMA) with 1% Tween 80. (2005)<sup>(23)</sup>.

- Sugar Fermentation Test : a set of sugars consists of Glucose, Maltose, Sucrose, Trehalose, Galactose, D-xylose and Lactose, which were used for identification and differentiation between *Candida* spp., the test was done by inoculating tubes containing fermentation media and 2% sugar with part of the colony, shaking gently then incubated at 28-30C° for3 days. The positive result was recorded by changing the color of phenol red to yellow and production of CO2 gas bubbles in Durham tube <sup>(25)</sup>.

- Carbohydrate Assimilation Test: the test depends on the ability of different species of yeasts to grow in various sugar solutions (glucose, lactose, trehalos, raffinose, Maltose, Melibiose, Galactose, D-xylose and sucrose).

Carbohydrate assimilation medium was poured in Petri dishes and inoculated with *Candida* spp., then wells were made by cork borer in the inoculated plates, each well were filled with 2% sugar and incubated plates at  $30C^{\circ}$  for 2-4 days <sup>(25)</sup>.

2. Haemolysin Activity test: Determination of haemolysin activity was evaluated with a blood plate  $assay^{(26)}$ .

3. Protease Production test: The protease production was determined according to Aoki <sup>(27)</sup> using a test medium consisting of SDA plates containing bovine serum albumin (BSA).

## **Results and discussion:**

Out of 130 samples collected form patients undergoing chemotherapy 19(14.6%) fungal isolates were found .

Results of blood culture showed *candida* isolates were detected in 14(20.5%) blood culture while 5(8.1%) *candida* isolates were found sputum samples. No growth were detected in 54(79.5%) of blood culture and 57(91.9%) of sputum.

Table (1): candida spp. In blood and sputum specimens .

Culture results	Blood		Sputum		Total	
	No	%	No	%	No	%
Growth	14	20.5	5	8.1	19	14.6
No growth	54	79.5	31	91.9	111	85.4
Total	68	100	62	100	130	100

According to blood culture results the most frequently isolated fungal was *C.albicans* 8(57.2%) followed by *C.glabrata* 2(14.3%), *C.krusei* 3(21.4%), then *C.parasillosis* 1(7.1%) as illustrated in table (2).

Table (2): Number and percentage of identified fungal isolates from blood culture samples.

isolates if oill blood culture samples.				
Fungal isolates	Number of	%		
	isolates			
C. albicans	8	57.2		
C. krusei	2	14.3		
C. glabrata	3	21.4		
C. parasillosis	1	7.1		
Total	14	100		

The present study revealed that the rate of Candid isolates by used blood culture method in

immunocompromised patients (as shown in table (2). there are many studies which have been conducted on fungal isolates from bloodstream by used blood culture method<sup>(34)</sup> but in iraq less study for comparing this results to other study<sup>(28)</sup>.

Only one study in Iraq which have been conducted on candidemia was study by AL-Asehagi, (2012)<sup>(28)</sup> in Salah al din who found that C. albicans were isolated from (17.6%) blood, While that reported by Chen et al  $(2006)^{(29)}$  in Australia, who isolated (32.1%)Theoklis et al. (2005)<sup>(30)</sup> and Diekema et al. (2002)<sup>(31)</sup> who found that candida spp. were isolated from (32%) in USA, which were agree with our results. And were C.albicans (37.9%), were C.glabrata (13.8%) and were *C.parapsilosis* (11.2%)in study conducted by Hsiue et al(2009)<sup>(32)</sup> in Taiwan. while in study reported by Lynn et al (2002)<sup>(33)</sup> studied 50 blood samples collected from immunocompromised patients in Texas and the results showed that (76%) of samples were candida spp of which *c.albicans* and *C.glabrata* (24%) for each of them, were *C.glabrata* (18%) were *C.krusei*. Kovacicova et al  $(2001)^{(34)}$  at in the Slovak Republic was isolated (52.9%) were C.albicans, were (32.1%) were non C.albicans of prospective was completed on 140 patients with fungaemia.

these different results may be due to the difference of technique for blood culture as the sample size to the volume of media and daily inverting inoculated broth with blood sample. In addition, the method used in our study by BHI broth/agar as show in Fig(1).



Fig 1: Blood culture in BHI agar/broth media.

For sputum samples the results showed that C.albicans were most isolated 3(60%), followed by *C.glabrata* 2(40%), as illustrated in table (3).

Table (3): Fungal isolates from sputum culture samples.

Fungal isolates	Number of	%
	isolates	
C.albicans	3	60
C. glabrata	2	40
Total	5	100

our results for sputum culture showed the *C.albicans* were more (60%).

There were many studies which have been conducted on opportunistic fungi in immunocompromesid patients. One of these studies was by Yongabi *et*  $al.(2009)^{(36)}$  isolated 12 isolates of *C.albicans* from 98 sputum. which were agreement with our results ,and Jaffer (1998)^{(37)} in Babylon province reported that *C. albicans* (69.2%) was the most common isolate in a study of pulmonary fungal infection, less frequency are *C. tropicalis* (19.2%), *C. kefyr* (7.6%) and *C. krusei* (3%). In other study were 26 (27%) of *C.albicans* and non albicans spp. were 29(30%) <sup>(38)</sup>. The reason for these variations in all studies may be due to sample size, environment factors, nutrition requirements and virulence factors of this fungi <sup>(39)</sup>, It is not easy to determine the pathogenic role of fungal isolates from the respiratory tract, to differentiate between infections, colonization and

contamination <sup>(40)</sup>.

# 2. Identification of *Candida spp*. Isolates:

*Candida* was identified depending on the morphological features on culture medium, germ tube formation, pseudohypha formation and chlamydospores according ( $^{41, 42}$ ) showed table (4).

Candida spp.	C.albicans	C.krusei	C.glabrata	C.parasillosis
Color colony	white to	white to	white to cream	white to cream
	cream	cream		
Germ tube	+	_	_	_
Chlamy-dospore	+	_	_	_
Pseudo-hyphae	+	+	_	+

 Table (4): Morphology feature and some tests used for identification of Candida Spp.

(+) positive , (\_): negative.

The *C.albicans* isolates was showed positive result and the formation of germ tubes was seen as long tube like projections extending from the yeast cells (Fig 2A), there was no constriction at the point of attachment to the yeast cells. These tubular extensions represent an early stage in the formation of true hyphae. The germ tubes were formed within two hours of incubation and this is a unique diagnosis characteristic of *C. albicans* differentiates it from other fungi. This results agreement with Mohammed  $(2008)^{(43)}$ , when they mentioned that "All *C. albicans* strains were germ tube test (GTT) positive when tested directly from the colony, and all non-*albicans*  species were GTT negative when tested directly from the colony".

C.albicans produced Chlamydospore than other species. They were spherical, thick-wall, and usually produced on suppurating cells that occur along pseudohyphae or at the tip of hyphae (Fig 2B). Approximately 90% Isolates produced of when inoculated chlamydospore by Dalmau technique or streak technique on cornmeal-Tween 80 agar . Pseudohyphae and hyphae with clusters of blastospores are also produce on this agar fig (2C). C.parasillosis, C.krusei and C.glabrata dosents produce chlamydospores (44).



Fig(2): A: Germ Tube of C.albicans stained by gram stains (40X) B:Chlamydospores of C. albicans Cultured on CMA-tween 80 at 30°C staning by gram stain B: Microscopic morphology of *C. albicans* in mycelial phase with blastoconidia budding from the pseudohyphae staining by LPCB stain (40x).

The biochemical tests were performed according to the tests described by Ellis *etal.* $(2007)^{(45)}$ .

The isolates were identified by used urease test, sugar assimilation and sugar fermentation. The results of this tests as in table (5).

Candida spp	C.albica-ns	C.krusei	C.glabrata	C.parasillosis	
Urease	_	_	_	_	
	Ferr	mentation	tests		
Glucose	+	+	+	+	
Maltose	+		+	_	
Sucrose	_	_	_	_	
Trehalose	V	I	V	_	
Galactose	V	I		+	
D-xylose	+			+	
Lactose	_			_	
Assimilation tests					
Melibiose	_	_	_	+	
Raffinose	_	_	_	_	
Glucose	+	+	+	+	
Maltose	+	_	_	+	
Sucrose	V	I	I	+	
Trehalose	+	I	V	+	
Galactose	+	_	_	+	
D-xylose	+	_	_	+	
Lactose	_	_	_	_	

Table (5): Biochemical tests used for identification of *Candida* spp.

(+) positive , (\_): negative , (V): variable .

# **4.3.1** Screening of protease production fungi isolates.

In this study, the extracellular protease production of *Candida* spp., was detected in 8(72.8%) of tested *C.albicans*, 5(40%)of *C.glabrata* and 1(100%) *C.parasilliosis*, while *C.krusei* were unable to protease production showed table (6).

Table (6): Protease Production by fungal isolates.

Fungal isolates	Number	Protease	
	isolates	No %	
C.albicans	11	8	72.8
C.glabrata	5	2	40
C.krusei	2	0	0
C.parasillosis	1	1	100

Result of present study agree with where De Bernardis <sup>(46)</sup> reported high *in vitro* protease activity in all *C. parapsilosis* strains isolated in patients with also Kanatrcioglu and Yucel <sup>(12)</sup> reported *in vitro* protease production in a majority of *C. albicans, C. kefyr, C. lipolytica, C. parapsilosis* and *C. tropicalis* clinical isolates, while *C. glabrata, C. guillermondii, C. krusei, C. lusitaniae* and *C. rugosa* were unable to protease positive. While Yamamoto (1992)<sup>(47)</sup> discovered that the majority of *C. tropicalis* and *C. parapsilosis* isolates had proteolytic activity while none of the tested *C.glabrata* strains secreted the enzyme.



Fig (4): Protease activity A: C.albicans. B: C.glaabrat

# **3.** Screening of hemolysis production by fungi isolates:

our results indicated that 11(100%) of C.albicans 4(80%) of C.glabrarta, 1(50%) of C.krusei and (100%) of *C.parasillosis* isolates were able hemolysin production showed table (7), These findings agree with Ramesh et al.<sup>(49)</sup> who compared the hemolytic activity of 50 Candida strains isolated from patients with HIV and 10 Candida strains isolated from immunocompetent patients. All strains produced hemolysis, but haemolytic activity was significantly higher for C.albicans strains isolated from HIV patients when compared to those isolated from immunocompetent patients. And Rossoni et al.<sup>(50)</sup> reported that *C.albicance* (100%) produced hemolysins, C.glabrarta and C.krusei each of all, (40%) of C.parasillosis isolates were hemolysin.

Tungui isolutesi				
Fungal isolates	Number	Hemolysis		
	isolates	No	%	
C.albicans	11	11	100	
C.glabrata	5	4	80	
C.krusei	2	1	50	
C.parasillosis	1	1	100	

 Table (7): hemolysin Production some by some fungal isolates.

Luo *et al.* (2001)<sup>(51)</sup> observed that species of *Candida* are capable of producing one or more types of hemolysins *in vitro* and that species differ in the production of these activities.

These differences are possible to be due to the environmental conditions, the source of isolate and the detection method that was used in the detected of the enzyme.

### Reference

1. William J, Steinbach F. (2003) Status of medical mycology education. J Med Mycol;41, 457\_467.

2. Cohen R, Roth F J, Delgado E, Ahearn D G, Kalser M H. Fungal flora of the normal human small and large intestine. N Engl J Med 1969 ; 280(12):638-41.

3. Rinaldi MG. Biology and Pathogenicity of Candida Species. In: Bodey GP, ed. Candidiasis Pathogenesis, Diagnosis, and Treatment. 2 ed. New York: Raven Press, 1993:1-20.

4. Pfaller, M.A. and Diekema. D.J. (2007) The epidemiology of invasive candidiasis: a persistent public health problem. Clin. Microbiol. Rev. 20:133-163.

5. Jarvis WR. Epidemiology of nosocomial fungal infections, with emphasis on Candida species. Clin Infect Dis. 1995. 20(6):1526-30.

6. Hilmar W, Tammy B, Sandra M T, Harald S, Richard P W, Michael B E. Nosocomial bloodstream infections in US hospitals: analysis of 24,179 cases from a prospective nationwide surveillance study. Clin Infect Dis 2004 Aug 1;39(3):309-17.

7. Walsh T J, Chanock S J. Diagnosis of invasive fungal infections: advances

8. Tomee JFCH, Kauffman H.F. (2000) Putative virulence factors of Aspergillus fumigatus. Clin Exp Allergy, 30:476–484.

9. Mio T, Yabe T, Sudoh M. et al. Role of three chitin synthase genes in the growth of Candida albicans. J Bacteriol. 1996;178:2416–2419.

10. Casadevall A, Pirofski LA. (2001) Host-pathogen interactions: the attributes of virulence. J Infect Dis 184:337–344.

11. Odds, F.C. (1985) Candida albicans proteinase as a virulence factor in the pathogenesis of Candida infections. Zbl. Bakt. Hyg. Ser. A., 260: 539-542.

12. Deng A, WU J, Zhang Y, Zhang G and Wen T. (2010) Purification and characterization of a surfactant-stable high-alkaline protease from Bacillus sp. , J Technol. 101: 7100-7116.



Fig (5): hemolysin activity of *C.albicans* isolate fraction of hemolysis in sheep blood agar at 37 °C.

In conclusion, according to results of this study *C.albicans* are the most common specie of blood among the total *Candida* infections, and most *candida* species isolates have ability to produce significant amount protease, hemolysin.

13. Hube B, Ruchel R, Monod M, Sanglard D, Odds FC (1998) Functional aspects of secreted Candida proteinases. Adv Exp Med Biol 436:339–344.

14. Borg, M. and Ruchel, R. (1988) Expression of Extracellular Acid Proteinase by Proteolytic Candida s during Experimental Infection of Oral Mucosa. Infection and Immunity, 56, 626-631.

15. Odds, F.C. (1985) Candida albicans proteinase as a virulence factor in the pathogenesis of Candida infections. J. Med Infect, 260: 539-542.

16. Ruchel, R. (1986) Cleavage of immunoglobulin by pathogenic yeast of the genus Candida. Microbiol. Sci., 3: 316-319.

17. Yuan, L. & G. T. Cole. 1987. Isolation and characterization of an extracellular proteinase of Coccidioides immitis. Infect Immun 55: 1970-1978. 22.

18. Kanatrcioglu SA, Yucel A. Phospholipase and protease activities in clinical *Candida* isolates with reference to the source of strains. Mycoses 2002;45: 169-5.

19. Monod M, Zepelin MB. (2002) Secreted proteinases and other virulence mechanisms of Candida albicans. Chem Immunol 81:114–128.

20. Naglik JR, Challacombe SJ, Hube B. (2003) Candia albicans secreted aspartyl proteinases in virulence and pathogenesis. Microbiol Mol Biol Rev 67:400–428

21. Glenn D Roberts and John A Washington. Detection of Fungi in Blood Cultures. J ClinMicrobiol. 1975.:309-310

22. Binesh, L. Y. and Kalyani, M. 2011.Phenotypic Characterization of Candida Species and Their Antifungal Susceptibility from a Tertiary Care Centre, J Pharm Bio Sci, 11: 12.

23. Segal, E., and Elad, D. (2005) Candidiasis. In Topley and Wilson s Medical Mycology. 10th edn. Edward Arnold Publishers. 579-623.

24. Atlas R. M., Williams J. F., Huntington M. K. (1995). *Legionella* contamination of dental-unit waters. *Appl. Environ. Microbiol.* 61: 1208–1213.

25. Forbes. S.h., hogg, J. T., Buchanan, F.C., Crawford. A. M. and Allendorf . F.W. (1995).microsatellite evolution in coneneric ma,,als: domestic and bighorn sheep. Mo. Boil. Evol. 12: 1106-1113.

26.Manns JM, Mosser DM, Buckley HR. (1994) Production of a Hemolytic Factor by Candida albicans. Infect Immun. 62;5154-6.

27. Aoki S, Ito-Kuwa S, Nakamura Y, Masuhara T. (1990) Comparative pathogenicity of wild-type strains and respiratory mutants of Candida albicans in mice. Bio Sci. 273: 332-343.

28. AL-Asehagi A. S. H (2012). Candidemia in Patients Undergoing Chemotherapy , College of Medicine University of Tikrit.

29. Chen S, Slavin M, Nguyen Q, et al. (2006)Active surveillance for candidemia, Australia. J Emerg Infect Dis. 12(10):1508-16.

30. Theoklis E Z, Jesse A.(2005)The Epidemiology and Attributable Outcomes of Candidemia in Adults and Children Hospitalized in the United States: A Propensity Analysis. Philadelphia. J Clin Infect Dis. 41:1232–9.

31. Diekema DJ, Messer SA, Hollis RJ, et al. (2003) Activities of caspofungin, itraconazole, posacoanzole, ravuconazole, voriconazole, and amphotericin B against 448 recent clinical isolates of filamentous fungi. J Clin Microbiol. 41:3623–6.

32. Hsiue h. c, y.T Huany ,Y. L. kuo, T. C. chang (2009). rapid identification of fungal pathogens in positive blood culture using oligonucleotide array hybridization .Clin Microbiol Infect 2010;16:493-500.

33. Lynn, A.G., MacDonald, W.A., Joanis, V., Lloyd, V.K. (2002). Developmental timing and maintenance of genomic imprinting and position-effect variegation in Drosophila. A. Dros. Res. Conf. 43 : 301A.

34. Kovacicova, G; Hanzen, J; Pisarcikova, M; Sejnova, D; Horn, J; Babela, R; Svetlansky, I; Lovaszova, M; Gogova, M. & Krcmery, V. (2001). Nosocomial fungemia due to amphotericin Bresistant Candida spp. in three pediatric patients after previous neurosurgery for brain tumors. *J Infect Chemother* 7, 45-38.

35. Forbes BN, Sahm DF, Weissfield AS. (2002) Bailey and Scott's Diagnostic Microbiolog, 9th ed. Chapter 53 M Mosby. St. Louis London Philadelphia Sydney Toronto, 280:724-785.

36. Yongabi, K.A. W.F.Mbacham, K.K.Nubia and R.M.Singh (2009) Yeast strains isolated from HIV-seropositive patients in Cameroon and their sensitivity to extracts of eight medicinal plants. African Journal of Microbiology Research, 3 (4), 133-136.

37. Jaffer WN. (1998) Pulmonary fungal infections in Babylion province. Ph D. Sc thesis, College of Sciences University of Babylon.

38. Bharathi B, Siva Sankar S, Swamidoss Danial (2010). Incidence of bacterial and fungal coinfections

in some HIV infected Indian population. Indian J. Biotechnol., 3(2): 199.

39. Cooke NS, Feighery C, Armstrong DK, Walsh M, Dempsey S. (2009) Cutaneous Fusarium solani infection in childhood acute lymphoblastic leukaemia. Clin Exp Dermatol 34:e117–e119.

40. Paradowski LJ. Saprophytic fungal infections and lung transplantation – revisited. J Heart lung Transpl. 1997; 16: 524-531

41. Osmanagaoglu, O.; Altinlar, N.; Sacilik, S.C.; Cokmus, C. and Akin, A. (2000). Identification of different Candida species isolated in various hospitals in Ankara by fungichrom test kit and their differentiation by SDS-PAGE. Turk. J. Med. Sci. 30: 355-358.

42. Bhavan, P.S.; Rajkumar, R.; Radhakrishnan, S.; Seenivasan, C. and Kannan, S. (2010). Culture and identification of Candida Albicans from vaginal ulcer and separation of enolase on SDS-PAGE. Inter. J. Bio. 2; (1): 84-93.

43. Mohammed N. A.(2008) Detection of Candida spp. and other pathogens responsible for vulvovaginitis in women with contraceptive methods. College of Science / University of Baghdad.

44. Pfaller MA, Diekema DJ. Twelve years of fluconazole in clinical practice: global trends in species distribution and fluconazole susceptibility of bloodstream isolates of Candida. *Clin Microbiol Infect* 2004 Mar;1:11-23.

45. Ellis, d., davis, s., alexiou, h., A., Bozza, S., Gaziano, R., Mosci, P(2007). Descriptions of medical fungi, second edition, underdale, south Australia: nexus print solutions.

46. De Bernardis F, Mondello F, San Milan R, Ponton J, Cassone A. (1999) Biotyping and virulence properties of skin isolates of Candida parapsilosis. J Clin Microbiol, 37:3481-86.

47. Yamamoto T, Nohara K, Uchida K, Yamaguchi K.Purification (1992). characterization of secretory proteinase of Candida albicans. Microbiol Immuno, 36: 637-41.

48. Garver KI, Muriana PM (1993). Detection, Identification and characterization of bacteriocin producing lactic acid bacteria from retail food products. Int. J. Food Microbiol. 19: 241-258.

49. Ramesh N, Priyadharsini M, Sumathi CS, Balasubramanian V, Hemapriya J, Kannan R. (2011) Virulence factors and anti-fungal sensitivity pattern of Candida sp. isolated from HIV and TB patients. Indian J Microbiol. 51(3):273-8.

50. Rossoni RD, Barbosa JO, Vilela SFG, Jorge AOC, Junqueira JC . (2013) Comparison of the hemolytic activity between C. albicans and non-albicans Candida species, J Clin Microbiol, 27(6):484-9 485.

51. Luo, G., Samaranayake, L. P. & Yau, J. Y. (2001). Candida species exhibit differential in vitro hemolytic activities. J Clin Microbiol 39, 2971–2974.

عزل وتشخيص انواع المبيضات لدى المرضى المثبطين مناعيا والتحري عن بعض عوامل الضراوة

ميلاد عدنان مزهر<sup>1</sup> ، وعد محمود رؤوف<sup>2</sup> ، خليل ابراهيم بندر<sup>3</sup> <sup>1</sup>قسم علوم الحياة ، كلية التربية للعلوم الصرفة ، جامعة تكريت ، تكريت ، العراق <sup>2</sup>كلية الصيدلة ، جامعة تكريت ، تكريت ، العراق <sup>3</sup>قسم علوم الحياة ، كلية العلوم ، جامعة تكريت ، تكريت ، العراق

#### الملخص

تم جمع 130 عينة دم وقشع من المرضى الخاضعين للعلاج الكيمياوي الراقدين والمراجعين لمستشفى تكريت التعليمي من كلا الجنسين من تتراوح اعمارهم ( 11–83 ) سنة ، تم جمع 68 عينة دم و62 عينه قشع ضمن هذه الدراسة.

بلغت نسبة الإصابة بالفطريات (%20.5% عزله من مجرى الدم و (%8.1% عزلة من القشع ، كانت اعلى نسبة اصابات من مجرى الدم هي C.parasillosis بنسبة (%57.8% وكانت C.glabrata بنسبه (%21.4 و 321.4% بنسبة (%21.4) و C.parasillosis و C.parasillosis بنسبة (%21.4 بنسبة (%17.1) بينما اكثر العزلات من القشع كانت لل C.albicans بنسبة 3(%60) و كانت C.glabrata بنسبه (%40) حي تم تشخيص العزلات وفقا لخواصها الشكلية والمز رعية و الكيموحيوية .