

Epidermal Growth factor in human urine as promotor for the growth of *Leishmania* sp. *In vitro*.

Nuha S. Al-Bayatii¹, Fatima Sh. Al-Naserii², Jaladet M.S. Jubrael³

¹ Department of Clinical Laboratory Sciences, College of Pharmacy, Tikrit University, Tikrit, Iraq

² Department of Biology, College of Sciences, Tikrit University, Tikrit, Iraq

³ Scientific Res. Center, College of Sciences, University of Duhok, Duhok, Iraq

Abstract

Leishmania parasites are the causal agents of leishmaniasis, a group of protozoan diseases transmitted to mammals, including human beings, by phlebotomine sandflies. In culture media (at 25-28°C, pH = 7.2-7.4), leishmania parasites develop as motile promastigotes similar to those found in the sand fly midgut. Traditionally media available do not meet the requirement for the bulk cultivation of Leishmania parasites, it requires fetal calf serum (FCS), that is very expensive and not easily available in the market. A number of studies have shown that the addition of 5-10% normal human urine stimulates growth, leading to more rapid multiplication and a higher concentration of parasites. Urine from patients with bladder cancer were used in this study to determine the effect of Epidermal growth factor (which is increased in level in this type of urine), and our study showed that proliferation indexes were significantly increased in the culture media supplemented with human urine from patients with bladder cancer, we undertook a detailed study of such an effect in old world *Leishmania* isolates causing cutaneous Leishmaniasis. We also found that urine with high percentage of EGF. Could be used as an alternative of fetal calf serum.

Key words: *Leishmania*, Epidermal growth factor, bladder cancer.

Introduction

Leishmania parasites are the causal agents of leishmaniasis, a group of protozoan diseases transmitted to mammals, including human beings, by phlebotomine sandflies [1]. In man and other hosts it occurs as a non-flagellar amastigote form, while in culture and gut of sandflies the flagellar or the promastigote form is seen [2]. Cutaneous leishmaniasis caused by *L. tropica* produces painless, frequently multiple, dry ulcers of the skin, The incubation period is usually 2–8 months [3]. Cutaneous leishmaniasis caused by *L. major* is, like other forms of cutaneous leishmaniasis, painless when the lesions are uncomplicated. The lesions are often severely inflamed and ulcerated and heal within 2–8 months [4]. Several different forms of promastigote have been described in sand fly infections [5]. A variety of culture medias designed for *in vitro* maintenance and bulk cultivations of *Leishmania* promastigotes require inclusion of 10-30% heat inactivated foetal bovine serum (HIFBS) to support successful growth [6]. Without HIFBS, these culture medias simply fail to support growth of *Leishmania* promastigotes and the culture dies off. Foetal bovine serum is not only the most expensive ingredient of these culture media but is also very difficult to obtain in many parts of the world where *Leishmania* is endemic [7]. Several studies have shown the stimulatory effect of human urine on *Leishmania* promastigotes, When supplemented with human urine, [8]. Epidermal growth factor or EGF is a growth factor that stimulates cell growth, proliferation, and differentiation by binding to its receptor EGFR. Human EGF is a 6045-Da protein, with 53 amino acid residues and three intramolecular disulfide bonds [9]. Epidermal growth factor can be found in human platelets, macrophages, urine, saliva, milk, and plasma. Increased activity of the receptor

for EGF has been observed in certain types of cancer, often correlated with mutations in the receptor and abnormal function such as constitutive receptor signaling independent of the levels of EGF or of binding of EGF [10]. Addition of EGF to was found to boost significantly proliferation of the bloodstream stage [7].

Aim of the study: To evaluate a new media for primary isolation of leishmanial different stages and detecting these stages of life cycle *in vitro* culture by using urine from patients with bladder cancer.

Material and Method:

1-Sampling: samples were taken from the lesions of 50 patients attending Tikrit teaching hospital in Tikrit governorate and diagnosed clinically by special dermatologist in dermatological department in the hospital from January -May 2013. All patients underwent lesion aspiration [11].

2- Preparation of media:

Lesion aspirate obtained previously were cultured into three groups of media, the first group tubes containing NNN media, It's composed of two phases, one is a solid phase and the other is liquid phase, this media used for cultivation and continuation of promastigotes stage of *leishmania* and used for the first time by Nove and Mac Neal 1904. (12,13) The second group were tubes containing RPMI 1640 (Roswell park memorial institute medium) (Sigma, St. Louis), L-glutamine (Sigma) medium supplemented with 10% FCS (prepared directly from blood of cow fetus, after separation in centrifuge then serum collected and inactivated in 50°C for 30min) and penicillin 1000U/ml and 0.3mg/ml streptomycin and nystatin 250 U/ml were added to avoid contamination [14]. The third group of tubes containing a semi solid medium (6) and a Fresh human Urine was obtained from Patients with bladder

cancer (attended to Tikrit teaching hospital), and made sterile by passing through 0.22 μ M filter paper. then the urine added in the medium tubes in percentage 5% of media and inoculated with the lesion aspirate (0.5ml).

All tubes were incubated at 25°C for 21 days and checked every two days by taking samples from each test tubes and by direct smear examination or smear doing by using Giemsa staining then re checked every two days until starting of growth. Parasites were counted with help of hemocytometer (WBC counting chamber) slide with a 40 \times objective of light microscopy, calculation done by the following equation:

Total number of promastigotes in ml = the number of promastigotes in 64 small square of haemocytometer $\times 25 \times$ dilution degree $\times 10^3$.

Result:

1-Direct smear: as shown in Figure 1, direct smear shows a different stages forms in (semisolid+urine) media.

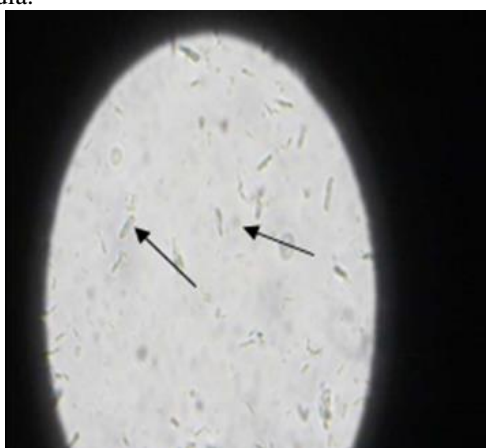


Figure 1: show different stages of *leishmania sp.* growth (direct smear)

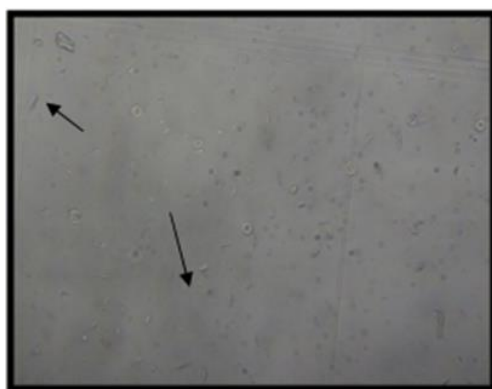


figure 2: show *leishmania sp.* Counting with WBC. counting chamber slide

2- Counting:

Figure 2, Shows (WBC counting chamber) slide for counting and numerous, different forms of *leishmani* promastigotes seen under the 40 \times objective of light microscope

3- Growth Rate: As shown in table (1): the mean growth rate of promastigote growth in NNN media, in which the recovery time was at eight day from

inoculum time, peak reached at fourteenth day from inoculation time then decline occurred for four days from the peak till reach to zero in number at eighteenth days of inoculation.

Table (1): Growth mean of *leishmania sp.* in NNN media

Days	Statistical analysis		
	Mean of growth $\times 10^6$	Standard deviation	\pm S.E.
8	0.53	1.72318	0.27245
10	0.91675	2.05565	0.32502
12	1.37	2.65515	0.419816
14	1.455	2.99692	0.473855
16	0.99125	2.44005	0.38580
18	0	0	0

While in table (2): shows the mean growth rate of promastigote growth in RPMI 1640 media with (FCS), in which the recovery time was at sixth day from inoculum time, peak reached at sixteenth day from inoculation time then decline occurred after six days later from the peak rate till reach to zero in number at twenty two days of inoculation.

Table (2): Growth means of *leishmania sp.* in RPMI 1640 + FCS. media.

Days	Statistical analysis		
	Mean of growth $\times 10^6$	St standard deviation	\pm S.E.
6	0.75525	1.17787	0.1862
8	1.21475	1.67081	0.2641
10	1.76275	1.97455	0.3122
12	2.42525	2.25508	0.3565
14	3.09625	2.77361	0.4385
16	3.19171	3.25526	0.5147
18	2.0395	2.20109	0.3480
20	0.897	1.02404	0.1619
22	0	0	0

Table (3), the recovery time was after four days of inoculation in semi-solid media supplemented with urine from patient with bladder cancer and the peak was in the fourteenth day of inoculation and decline after seven days after that till reach to zero in number at twenty two days of inoculation.

Table (3): Growth mean of *leishmania sp.* in Semisolid + Urine media.

Days	Statistical analysis		
	Mean of growth $\times 10^6$	standard deviation	\pm S.E.
4	1.3695	1.32909	0.2101
6	2.0485	1.5951	0.2522
8	2.841	2.2392	0.354
10	3.6025	2.4638	0.3895
12	4.5577	2.85089	0.4507
14	5.592	3.23677	0.51177
16	5.53	3.317	0.5246
18	3.268	2.5034	0.3958
21	0.74525	1.88277	0.29769
22	0	0	0

-Figure 3, shows the differences in the three different medias (NNN, RPMI 1640, Semi-solid with urine), and as show the peak rate was in Semi-solid media

supplemented with urine from patients with bladder cancer, followed by RPMI 1640 media .and the least growth were on NNN media.

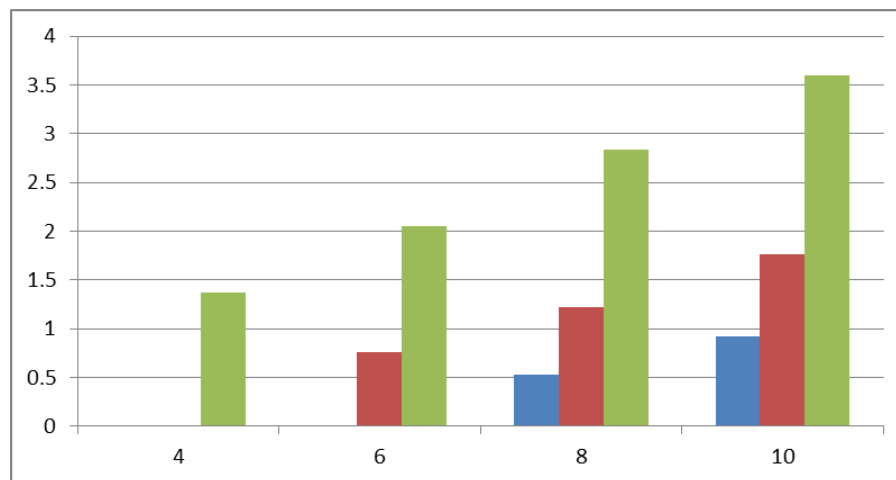


Figure (3): Growth of Leishmaniapromastigotes in different types of media :NNN(Blue), RPMI+FCS(Red), Semi solid media +Urine form bladder cancer patients(Green)

Discussion:

A variety of culture Medias designed for *in vitro* maintenance and bulk cultivations of *Leishmania* promastigotes require inclusion of 10-30% heat inactivated fetal bovine serum (HIFBS) to support successful growth. Fetal bovine serum is not only the most expensive ingredient of these culture media but is also very difficult to obtain in many parts of the world where *Leishmania* is endemic [6], table (2) of this study proves that that growth peak of isolated cutaneous leishmania was at sixteenth (3.19171×10^6) promastigotes/ml day of inoculation which is faster to that time of growth on ordinary NNN media in table (1) which shows the recovery time of promastigote was at eighth day and the peak of growth was at fourteenth day with (1.455×10^6) promastigotes/ml and longer to the time of result of Mobarak [15], who found that growth peak of isolated cutaneous leishmania was 2.8×10^6 at thirteen day of inoculation and Limoncue *et al.* [16] who use RPMI 1640 and found the peak of growth was 19×10^6 at ten day of inoculum and this could be due to different inoculum dose (as the inoculum dose was 10^5 cell/ml) and difference of strain of this parasite or different culturing environment. Also this table shows that the recovery time was at sixth day after inoculum and this result agrees with Andrea *et al.* [17] who found the median of isolation time in RPMI 1640 with FCS was six day, and faster to Mobarak [6], study who found that the recovery time of leishmania parasite was eight day this could be due to different culture method used. It was observed in table (3), the recovery time was in fourth day and the peak rate was (5.592×10^6) promastigotes /ml after fourteenth days of inoculum and this result proves that human urine from patient with bladder cancer promotes the growth of leishmanial promastigote

forms *in vitro*, compared with RPMI 1640 media supplemented with fetal calf serum and NNN media. The major constituent of the normal urine includes proteins, amino acids, urea, uric acid, succinate, alanine, citrate, creatine, creatinine, dimethylamine, Formate, glycine, hippurate, histidine, indoxyl sulfate, lactate, N-acetyl groups from glycoproteins, phenylalanine, taurine, trimethylamine N-oxide, 3-hydroxybutyrate, acetate [18,19]. Xanthine [20] and many studies have shown that the addition of 1-5% human urine stimulates growth, leading to more rapid multiplication and a higher concentration of parasites at the stationary phase [8]. While in other studies which found that the constituent of urine could be different in patients according to the case history for instance, hyperglycemic patients, hyperproteinemia, however the presence of glucose at concentration of 100-200 mg/24hr. and amino acids at concentration 1.5g/24hr. can play a major role as growth promoting factor for leishmania which can use several carbohydrate as respiratory substrate including glucose, fructose, mannose [21]. Several studies demonstrated the role of amino acids as primary growth substrate, its reported that promastigotes take up proline at the rate 5 times faster than that of glucose [22]. Allahverdiyeva *et al.* demonstrated that changes in proliferation and infectivity of leishmania parasites may be affected by components found in urine. Leishmania parasites have a digenetic life cycle. They first differentiate and increase in size as procyclic promastigotes within the gut of *Phlebotomus* spp. sand flies. They then exit the cell cycle and differentiate as metacyclic promastigotes (infective form). Growth curves of procyclic and metacyclic forms of promastigotes can change in *in vivo* culture conditions, and some differences also accrued during

life cycles of parasites [8,18]. Other components of urine are the epidermal growth factor amitogenic polypeptide produced by many cell types and made in large amounts by some tumors. It promotes growth and differentiation, is essential in embryogenesis, and is also important in wound healing. It has been found to be part of a family of compounds that includes also transforming growth factor [23]. Hide *et al.* have shown the presence of an epidermal growth factor (EGF) receptor in *T. brucei*, raising the possibility that this and other cytokine or cytokine-like substances might regulate parasite growth in the host. Addition of EGF to MEM supplemented with cysteine and BCS was found to boost significantly proliferation of the bloodstream stages [24]. The optimal range for EGF was 20 to 200 nM, many times higher than the reported concentration of EGF in human blood (0.02 to 0.3 nM). According to Sternberg and McGuigan, EGF was probably acting as a mitogen [25]. The disparity between the mitogenic stimulatory effect and the bloodstream concentration of EGF suggested that there may be other low-molecular-weight compounds in blood that stimulate proliferation of bloodstream stages in early stages of infection of the mammalian host [24].

References:

- [1] Reithinger, R. Dujardin, J.C. Louzir, H. Pirmez, C. (2007). Cutaneous leishmaniasis. The lancet. (7).
- [2] UIBari, A. Ber Rahman, S. (2008). Cutaneous leishmaniasis: an overview of parasitology and host-parasite-vector inter relationship. Pakistan Association of Dermatologists J. 18: 42-48.
- [3] WHO Expert Committee on the Control of Leishmaniases. (2010). Control of the Leishmaniasis. Report of a meeting of the Geneva, 22–26.
- [4] Khan, S.J. and Muneeb, S. (2012). Cutaneous leishmaniasis in Pakistan. Dermatology Online Journal 11 (1): 4.
- [5] Bates, A. P. (2007). Transmission of *Leishmania* metacyclic promastigotes by phlebotomine sand flies. Int J Parasitol, 37(10-3): 1097–1106.
- [6] Iqbal, J. Jamshid, M. Ahmed, B. Bukhari, I. Bashir, S. Yasinzai, MM. (2006). Some studies on human urine as promoter for the growth of leishmania in vitro. Pak J Pharm Sci.; 19(2):152-5.
- [7] Schuster, F.L. and Sullivan, J.J. (2002). Cultivation of Clinically Significant Hemoflagellates. Clin. Microbiol. Rev.; 15 (3) 374-389.
- [8] Allahverdiyev, A.M. Bagirova, M. Elcicek, S. and Oztel, O.N. (2011). Effect of Human Urine on Cell Cycle and Infectivity of *Leishmania* Species Promastigotes *in Vitro*. Am. J. Trop. Med. Hyg., 85(4): 639–643.
- [9] Callegari, C. Laborde, N.P. Buenaflor, C.G. Brasel, J.A. and Fisher, D.A. (1988). The source of urinary epidermal growth factor in humans. European Journal of Applied Physiology and Occupational Physiology. 58(1):26-31.
- [10] Konturek, J.W. Bielaski, W. Konturek, S.J. Bogdal, J. and Oleksy, J. (1989). Distribution and release of epidermal growth factor in man. Gut, 30, 1194-1200.
- [11] Evans, D. (1989). Handbook on isolation, characterization, and cryopreservation of *Leishmania*. Special programme for research and training in tropical Diseases. WHO.
- [12] Kagan, I. G., and Norman, L. (1970). In "Manual of clinical microbiology" AM. Soc. Microbial, Washington, 453-486.
- [13] Dawson, R. M. C., Elliot, D. C., Elliot, W.H., and Jons, K. M. (1969). Data for biochemical research 2ed., Clarendonpress, Oxford, 508.
- [14] Mobarak, H.A. (2008). Isolation and Cultivation of Cutaneous *Leishmania* parasite by using Different Cultures Media. A thesis Submitted to the committee of postgraduate studies University of Kufa /College of the Medicine. IRAQ.
- [15] Mobarak, H.A. Tarish, H.R. Al Masudi, H. (2011). Isolation of cutaneous leishmania parasite by using RPMI1640 and Schneider drosophila media Kufa Med. Journal. 14(1):297-300.
- [16] Limoncu, M.E. Ozbilgin, A. Balcioglu, I.C. Ozbek, Y. (2004) Evaluation of three new culture media for the cultivation and isolation of *Leishmania* parasites. J Basic Microbiol.; 44(3):197-202.
- [17] Andrea, K., Bogylel, C. M., Diego, E., Jorge, A., and Vanessa, A. (2007). Evaluation of microculture method for isolation of leishmania parasite from Cutaneous leishmania of patient in Peru. J. Clin. Microbiol. 45: 3680-3684.

- [18] Erzaeg, Z.S., Al. Taie, A.A., (2013). Najim, W.S. Morphologic forms of Leishmaniapromastigotes in a new media .Tikrit J. Med.sci.
- [19] Pasihogios, N.G., Gazi, I.F., Elisaf, M.S. (2008). Gender –related and age-related urinalysis of healthy subjects by NMR-based metabonomics. NMR Biomed;21:195-207.
- [20] Warburg, A. Gelman, S. and Deutsch, J. (2008).Xanthine in urine stimulates growth of Leishmaniapromastigotes in vitro. Journal of Medical Microbiology;57(1):136-138.
- [21] Darling, T.N. Davis, D.G. London, R.G. and Blum, J.J. (1987).Products of Leishmanibraziliensis glucose catabolism: release of D-lactate and, under anaerobic conditions, glycerol. Proc Natl Acad Sci U S A ; 84(20): 7129–7133.
- [22] Vieira, L.L, Cabantchik, Z.I.(1995).Amino acid uptake and intracellular accumulation in Leishmania major promastigotes are largely determined by an pump generated membrane potential. Mol BiochemParasitol.; 75(1):15-23.
- [23] en.wikipedia.org/wiki/Epidermal_growth_factor
- [24] Hide, G.A. Gray, C. M. Harrison, and Tait, A. (1989). Identification of an epidermal growth factor receptor homologue in trypanosomes. Mol. Biochem. Parasitol. 36:51-60.
- [25] Sternberg, J. M., and. McGuigan, F.(1994). Trypanosomabrucei: mammalian epidermal growth factor promotes the growth of the African trypanosome bloodstream form. Exp. Parasitol. 78:422-424.
- [26]Armstrong, T. C., and Patterson, J.L.(1994). Cultivation of Leishmanibraziliensis in an economical serum-free medium containing human urine. J. Parasitol. 80:1030-1032.
- [27]Chow, N.H., Liu, H.S., Lee, E.I., Chan, S.H., Cheng, H.L., Tzai, T.S., Lin, J.S. (1997).Significance of urinary epidermal growth factor and its receptor expression in human bladder cancer. Anticancer Res.; 17 (2B): 1293-6.
- [28] Neal, D.E., Sharples, L. Smith, K. et al.(1990). The epidermal growth factor receptor and the prognosis of bladder cancer. Cancer;65:1619–25.

تحفيز نمو طفيلي الشمانيا بواسطة عامل نمو البشرة المأخوذ من ادرار الإنسان خارج الجسم الحي

نهى سليم البياتي¹، فاطمة شهاب الناصري²، جلادت محمد صالح جبرائيل³

¹ فرع العلوم المختبرية السريرية، كلية الصيدلة، جامعه تكريت، تكريت، العراق

² قسم علوم الحياة، كلية العلوم، جامعه تكريت، تكريت، العراق

³ مركز البحوث العلمية، كلية العلوم، جامعه دهوك، دهوك، العراق

الملخص

تعتبر طفيليات الشمانيا المسببات الرئيسية لداء الشمانيا. وهي مجموعة من الابتدائيات التي تنتقل للبائن ومن ضمنها الإنسان، بواسطة ذبابه الرمل. في الظروف الزرع عيه (بدرجه حرارة 25 - 28 م، ووسط قاعدي 7.2-7.4)، طفيلي الشمانيا يمكن ان يتطور الى طور إمامي السوط المتحرك مشابه لما موجود في داخل جسم الحشرة. استخدام الأوساط التقليدية لا يفي بالمتطلبات الغذائية لإنتاج أعداد كبيرة من الطفيليات حيث يتطلب أضافه مصل العجول حديثه الولادة والذي يكون عاده مكلف جدا وغير متوفر بشكل دائم. عدد من الدراسات أظهرت أضافه 5-10% من إدرار الإنسان يؤدي إلى تحفيز وزيادة النمو وتضاعف سريع وزيادة تركيز الطفيلي. في هذه الدراسة تم استخدام إدرار من مرضى سرطان المثانة لتحديد تأثير عامل نمو البشرة (والذي يزداد مستواه في هذا الإدرار) وأظهرت الدراسة زيادة في معدل النمو للطفيلي في الأوساط المزودة بهذا النوع من الإدرار كذلك وجد ان النسبة العالية من عامل نمو البشرة يمكن ان يكون بديلا لمصل العجول الحديثة الولادة .

الكلمات المفتاحية: الشمانيا، عامل نمو البشرة، سرطان المثانة .