# A Comparative study between the two interferon gamma releasing assays in the diagnosis of pulmonary tuberculosis

Asmaa Z. Al- Ghreer, Zainalabdeen A. Abdualla

Department of Microbiology, College of Medicine, University of Mosul, Iraq

#### Abstract:

**Objectives**: To examine and compare the diagnostic value of Interferon gamma releasing assay (IGRAs) (T-SPOT TB and Qunati FERON Gold in Tube) in the diagnosis of pulmonary tuberculosis.

**Materials and methods:** The study included 40 patients with active pulmonary tuberculosis. They were attendance of Respiratory Health Care Centers in Mosul City for the period from March to December 2013. From each patient a sputum sample was collected and processed for culture on Löwenstein-Jensen (L-J media). Twelve milliliters (ml) of blood from each patients was collected (10 ml collected in heparinised tube for T SPOT TB and 2 ml in QFT-GIT special tubes)

**Results:** From the total 40 patients with positive AFB staining 37 (92.5%) were culture positive. The specificity of both IGRAs were 100%. The sensitivity of T- SPOT TB test was 91.89%, while the QFT-GIT sensitivity was 86.49%. The positive predictive value of both IGRAs was 100%. The negative predictive value of T SPOT TB was 50%, while that of QFT-GIT was 37.5%. There was a moderate degree of agreement between the T -SPOT TB and QFT-GIT in (82.5%, k=0.54, 95%CI). The results of the two IGRAs are not affected by BCG vaccination status of TB patients.

**Conclusions**: The IGRAs could provide a supplementary information as part of diagnostic work-up for tuberculosis diagnosis but it is important to note that a negative IGRA does not rule out active TB. Moreover, and of the two IGRAs, QFT GIT is more convenient to adopted for diagnostic use.

Key words: Tuberculosis, T-SPOT TB, Quanti FERON Gold in tube.

## Introduction

With approximately 9 million patients annually, tuberculosis (TB) contributes significantly to worldwide mortality and morbidity, specially in lowincome countries. Despite lower TB mortality rates in high income countries, diagnosis and subsequent treatment of TB remains a health priority in order to prevent spread of disease and reduce the economic cost associated with patient care (1). However, TB control still relies on tests such as culture, smear microscopy and chest radiographs, despite their known limitations. Culture, the reference standard for active TB, is time consuming and often not available in resource poor settings. Smear microscopy, the most rapid and widely used TB test, is highly specific but has poor sensitivity (2, 3). More recently, two quantitative T-cell interferon-  $\gamma$  release assays (IGRAs), namely Quanti FERON-TB Gold In-tube (QFT-GIT; Cellestis, Carnegie, VIC, Australia) and T-SPOT TB test (Oxford Immunotec, Abingdon, United Kingdom), have been developed. These assays represent a long-awaited advancement in the field of TB diagnostics and are widely anticipated to replace the century-old tuberculin skin test (4). The IGRAs are based on the principle that upon exposure to Mycobacterium tuberculosis (MTB) in vitro, antigen specific T- cells present in the blood become activated and secret INF -  $\gamma$  (3,5, 6). The tests involve stimulation of blood T cells with the MTB antigen overnight and measurement of subsequent INF- $\gamma$  and the detection of INF -  $\gamma$  indicates TB infection (7). To avoid cross reactivity these tests use the antigen encoded in the region of determination 1 (RD1) (6,8). These antigens include the ESAT6 (early secreted antigen target -6) and CFP10 (culture

filtrate protein -10) along with TB 7.7 antigen (9, 10).

Since IGRAs cannot distinguish between latent TB infection and active TB, their use for the diagnosis of active disease has been extensively debated (4). The current work is aimed to examine and compare the diagnostic value of both commercially available IGRAs in the diagnosis of active pulmonary tuberculosis.

#### Subject, materials and methods

Forty patients apparently suffering from active pulmonary tuberculosis with positive sputum smear for acid fast bacilli were enrolled in this study. They were attendance of the Respiratory Health Care Centers in Mosul City. The collection of samples was carried out from March 2013 up to December 2013. A Questionnaire Form included information related to the patients and their diseases was completed.

## Sample collection and processing

An expectorated sputum sample was collected in sterile 50 ml container from each patient have positive direct smear for AFB. The smears stained using Ziehl Neelsen staining, the stains and procedure of staining were used as directed by Winn et al., 2006 (11). The patients were taught how to collect the sputum sample by coughing up the sputum deep from the lungs in a well ventilated areas. The sputum processed homogenization, sample was bv decontamination and concentration for culture on Löwenstein-Jensen (L-J) medium. Twelve ml of venous blood was drawn from each patient. Ten ml were obtained and transported in a heparinised tubes and used for T- SPOT TB test. Two ml were collected for QFT-GIT assay in special tubes provided by Cellestis, Australia, QIAGEN Company.

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The T-SPOT TB assay (Oxford, Immunotec, United Kingdom) is a simplified variant of the ELISPOT assay technique. The peripheral blood mononuclear cells (PBMCs) are separated from a whole blood sample and washed to remove any sources of background interfering signal. The PBMCs are then counted to achieve a standardized cell number is which was (250,000+ 50,000 PBMCs per well). The T -SPOT TB assay require 4 wells to be used for each sample. A nil control to identify non-specific cell activation using cell culture media incubated with the PBMCs, TB-specific antigens including Panel A (ESAT6) and Panel B (CFP10), and a positive control containing phytohaemagglutinin to confirm PBMCs functionality.

The T-SPOT was performed according to the manufactures training. The results for T -SPOT TB were interpreted by subtracting the spot count in the nil control well from the spot count in each of the two panels according to the following algorithms :

1. The test result is positive if the spots in Panel A minus spot in Nil and/or spots in Panel B minus Nil  $\geq 6$ .

2. The test result is negative if both ( spots Panel A minus spots in Nil) &( spots Panel B minus spots Nil)  $\leq 5$ .

3. The test results considered borderline where the highest of the Panel A or Panel B spot count is such that the (Panel minus Nil) spot count is 5,6 or 7 spots. The QFT-GIT test is performed in two stages according to the manufacturer's guiding instruction . First, whole blood was collected into each of the QFT –GIT blood collection tubes, the tubes were incubated at 37 Celsius as soon as possible, and within 16 hours of collection. Following a 16 to 24 hour incubation period, the tubes were centrifuged, the plasma is removed and stored at -20 Celsius in small aliquot

tubes. The amount of IFN- $\gamma$  (IU/ml) measured by ELISA, the test was considered positive when IFN- $\gamma$  response to the TB Antigen tube was  $\geq 0.35$  IU/ml.

### Statistical analysis

The data were analyzed statistically according to Dunn and Clack, 2009(12). The degree of agreement between T-SPOT TB test and QFT-GIT was calculated using Kappa test (k). Sensitivity, specificity, positive predictive value(PPV) and negative predictive value (NPV) were also calculated. **Results** 

The 40 patients included in the present study had an active pulmonary tuberculosis since they were all AFB –positive. Their age ranged between 13 to 65 years with mean  $\pm$  SD of  $35.9\pm$  13.65, of whom 24 (60%) were males and 16 (40%) were females. The medical history of these patients revealed all the 40 patients had a history of cough for more than one month and 38 (95%) had fever. Thirty three (82.5%) patients had history of night sweating, while 15(37.5%) had haemoptysis. Only 10 (25%) patients complained of loss of weight (Figure 1). The culture of the 40 AFB positive sputum samples showed that 37 (92.5%) had growth on L-J media and 3(7.5%) samples revealed no growth (culture negative).

The T -SPOT TB test gave positive result in 35 (87.5%) patients and all the T-SPOT TB positive patients were culture positive. A negative T-SPOT TB test was found in 5 (12.5%) patients. Two out of the 5 were culture positive and the other 3 were culture negative (Table1). There was a high statistical difference in the results of culture and T-SPOT TB test at *p*-value of < 0.001. The specificity of T-SPOT TB was 100%, while its sensitivity was 91.89 %. The PPV of this test was 100% and its NPV reached to 50 % (Table 1).

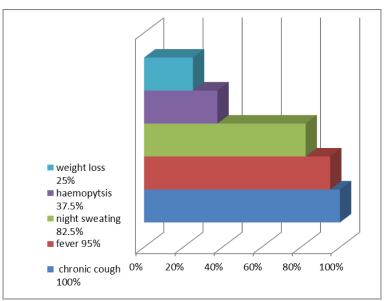


Figure 1: Clinical history in patients with tuberculosis

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The QFT-GIT immunoassay was positive in 32 (80%) and negative in 8(20%) patients. All the QFT-GIT positive patients were culture positive while, 5 out of the 8 patients with negative QFT-GIT were culture positive. The remaining 3 patients with negative QFT-GIT were culture negative (Table 5).

There was a high statistical difference between the results of mycobacterial culture and QFT-GIT at p-value <0.001. The specificity of QFT-GIT was 100%, while its sensitivity reached to 86.49%, and the test gave 100% PPV and 37.5% NPV(Table2).

TB patients	Culture positive	Culture negative	Total		
	n(37)	n(3)			
T-SPOT TB positive	35	0	35		
T-SPOT TB negative	2	3	5		
Total	37 3		40		
<i>p</i> -value	<0.001				
Specificity %	100				
Sensitivity%	91.89				
PPV %	100				
NPV %	50				

Table 1: T-SPOT	TB test and culture in tuberculosis p	oatients
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Table 2: QFT-GIT and culture results in patients with tuberculosis.

TB patients	Culture positive	Culture negative	Total		
	n(37)	n(3)			
QFT-GIT positive	32	0	32		
QFT-GIT negative	5	3	8		
Total	37	3	40		
<i>p</i> -value	<0.001				
Specificity %	100				
Sensitivity%	86.49				
PPV%	100				
NPV%	37.5				

The T-SPOT TB and QFT-GIT gave four patterns of concordance and discordance. The two concordance were the positive (T-SPOT TB +/QFT-GIT +) and the negative (T-SPOT TB -/QFT-GIT-). The two discordant types were T-SPOT TB +/ QFT-GIT- and T-SPOT TB-/QFT-GIT+.

The first pattern of concordance T-SPOT TB+/QFT-GIT+ was found in 31(77.5%) of patients. The other type of concordance T-SPOT TB-/QFT- GIT- was

observed in 4 (10%)patients. The discordant T- SPOT TB+/QFT- GIT- was detected in 4 (10%) patients, while the discordant pattern of T-SPOT TB-/QFT-GIT+ was found in only one patient (2.5%), Table 3. There was a moderate degree of agreement between T-SPOT TB and QFT- GIT as Cohens Kappa coefficient revealed a value of k=0.54 with agreement of 82.5 %.

Table 3 :	The concor	dance betw	een T-	Spot	TB and	QFT-G	HT in	patients	with	tuk	perculosis
			_					-		-	

TB patients	QFT- GIT positive	QFT -GIT negative	Total		
	no.(%)	no. (%)			
T-SPOT TB positive	31 (77.5)	4 (10%)	35		
T-SPOT TB negative	1 (2.5)	4 (10)	5		
Total 32 8 4					
Degree of agreement =82.5%					
Kappa = 0.54 (moderate agreement)					

The history of BCG vaccination in the patients included in the study was positive in14 (35%) and was negative in 26 (65%) patients. In the 35 positive T-SPOT TB patients, 11(31.43%) were BCG vaccinated and 24 (68.57%) were not vaccinated

while, in the 5 T- SPOT TB negative 3 (60%) vaccinated and 2 (40%) patients were non-vaccinated. There was no statistical difference between the state of BCG vaccination and the out- come of T-SPOT TB test results (Table 4).

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Vaccination status	T-SPOT TB positive	T-SPOT TB negative	Total		
	no.(%)	no.(%)			
BCG positive	11(31.43)	3(60)	14		
BCG negative	24 (68.57)	2(40)	26		
Total	35	5	40		
$X^{2} = 1.569$					
<i>p</i> -value 0.1 (Not significant)					

Table 4: T- SPOT TB test and BCG vaccination in patients with tuberculosis

In the 32 positive QFT-GIT, 10 (31.25%) gave positive history for BCG vaccination and 22 (68.75%) were non-vaccinated. The QFT-GIT negative patients were divided equally into BCG vaccinated and BCG non-vaccinated, 4 (50%) patients for each group. There was also, no statistical difference (p=0.25) between the results of QFT-GIT and BCG vaccination status in TB patients (Table 5).

 Table 5: QFT-GIT and BCG vaccination status in patients with tuberculosis

Vaccination status	QFT-GIT	QFT-GIT	Total			
	positive	negative				
BCG positive	10(31.25%)	4(50%)	14			
BCG negative	22 (68.75%)	4 (50%)	26			
Total	32	8	40			
$X^2 = 0.8$						
<i>p</i> -value 0.25 (Not significant)						

## Discussion

Several studies have tested the performance of the two IGRAs as diagnostic aid in mycobacterial diseases (13). The 100% specificity of T-SPOT TB test in detection of MTB denoted in the current study is in agreement with other studies (14, 15,16). The latter studies reported a specificity of T-SPOT TB test ranging from 98% to 100%. However, other works done by Simsek et al., 2010 (17) and Sester et al., 2011(18) reported lower specificity of 79% and 82% respectively. Furthermore, the sensitivity of T-SPOT TB test in the present study is 91.89%, which is slightly lower than that reported by Abdel Samea et al., 2013 (16), who recorded 100% sensitivity. Another study done by Simsek and Colleagues 2010 (17) reported a sensitivity of T- SPOT TB test of 51.4% which is lower than that of the current work. However, the sensitivity of T-SPOT TB in the present study is in agreement with the result of Biachi et al., 2009 (19).

The specificity of the QFT-GIT in the current study is 100%, which is reported also by several other studies done (15,16, 20,21). However, a lower specificity of 62.5 % has been recorded by Simsek *et al.*, 2010 (17). On the other hand, the sensitivity QFT-GIT in this study was 86.49% which is consistent with the results of Eddin and Monem, 2011(22). The discrepancies in the specificity and sensitivity of the IGRAs between different studies may be explained on the basis of the stages of TB disease (complicated or not), high versus low burden TB diseases areas and host factors, such as age, nutritional status, immunosupression and other comorbid conditions (e.g. diabetes mellitus).

In the current study, the two IGRAs have the same specificity but T-SPOT TB versus QFT-GIT is more

sensitive in detecting MTB infection. This was demonstrated by the higher positive results revealed by T-SPOT TB than QFT-GIT in patients with pulmonary TB. Whether such an increase in sensitivity could make this test clinically useful in the evaluation of active tuberculosis remains to be determined.

Furthermore, the T-SPOT TB test and QFT-GIT had a high PPV of 100%, which goes with the results of other studies (16, 20, 21). These studies reported a PPV that range from 85% to 100%. On the other hand, these studies also reported a high NPV from 80 % to100 % which disagrees with the results of the current study where the NPV was low for both T-SPOT TB test and QFT-GIT.

The concordance between T-SPOT TB test and QFT-GIT in this study revealed a moderate degree of agreement (k = 0.54) which goes with results demonstrated by Lee *et al.*, 2006 and Arend *et al.*, 2007 (22,24). In contrast to the present study, other studies ( (19, 25) compared the T-SPOT TB test and QFT-GIT in active TB patients obtained a high inter assay agreement of (83.2 % k = 0.66) rather than a moderate one.

The discrepancies between the results of the two test may be explained on the basis that the more sensitive test can detect the patients with affected immune system due to various immunosuppressive factors (disease, drug or old age). The immunosuppressive factors affect the CD4 memory T cells in reducing their ability to produce different cytokines including the INF-  $\gamma$  (26). Although, there is discrepancies in both tests but both are specific and sensitive enough to detect TB infection.

The two commercialized IGRAs are similar in terms of the antigens used (ESAT 6 and CFP10) and the

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incubation time (overnight or 16–24 h). The main differences between the two assays that QFT-GIT use a 3<sup>rd</sup> antigen (TB 7.7), the technique of IFN-  $\gamma$  detection is ELISA versus ELISPOT, and the specimens used are whole blood versus mononuclear cells. Of the two assays, QFT-GIT seems to be more convenient than T- SPOT, as it utilizes whole blood instead of mononuclear cells, beside ELISA is being more commonly used and simpler to perform than ELISPOT. In addition, the ELISPOT assay requires an expensive ELISPOT reader for accuracy, but is sensitive enough to detect single IFN-  $\gamma$  producing cells.

The evaluation of the results of BCG vaccination status and results of both T-SPOT TB test and QFT-GIT in the current study revealed that there is no statistical association between the two parameters in both TB patients. These results are consistent with several other studies done on active TB patients ( **References** 

**1.** Whiteworth HS, Scott M, Connell DW, Donge's B and Lalvani A (2013). IGRAs- the gateway to T-cell based TB diagnosis. **Methods**. **61(2013): 52-62**.

**2.** Ling DI, Pai M, Davids V., Brunet L, Lenders L., Meldau R, Calligaro G, Allwood B, Zyl-smit R, Peter J, Bateman E, Dawson R, Dheda K (2011). Are interferon gamma release assays useful for diagnosing active tuberculosis in a high burden setting. **Europ Respir J. 38 (3): 649-657.** 

**3.** Pai M., Kalantri S, Dheda K (2006). New tools and emerging technologies for the diagnosis of tuberculosis: part I latent tuberculosis. **Expert Rev Mol Diagn. 6(3):413-422.** 

**4.** Chee CB, Gan SH, Khinmar KW, Barkham TM, Koh CK Liang S and Wang YT (2008). Comparison of sensitivities of Two commercial Gamma Interferon Release assay for pulmonary tuberculosis. **J of Clinic Microb. 46(6): 1935-1940.** 

**5.** Tsioris SJ, Coetzee D, Toro PL, Austin J, Stein Z and Elsadr W (2006). Sensitivity analysis and potential uses of a novel Gamma Interferon release assay for the diagnosis of tuberculosis. J Clinic Microb. 44(8): 2844-2850.

6. Trajman A, Steffen RE and Menzies D (2013). Interferon gamma release assay versus tuberculine skin testing for the diagnosis of latent tuberculosis infection . Pulm Med. V. 2013, Article ID 601737, PP: 11.

7. Jacobs S, Warman A, Richardson R, Yacoub W, Lau A, Whittaker D, Cockburn S, Verma G, Boffa J, Tyrrell G, Kunimoto D, Manfreda J, Khassen D, Long D(2011). The tuberculin skin test is unreliable in school children BCG- vaccinated in infancy and at low risk of tuberculosis infection. **Pediatr Infect Dis** J. 30 (9):754-758.

**8.** Raja A (2004). Immunology of tuberculosis. **Indian J Medical Research** . **120** (1): **213-232.** 

**9.** Ramos JM, Robledano C, Masia M, Belda S, Paddle S, Rodriguez JC and Gutierrez F (2012). Contribution of interferon gamma release assays

## ISSN: 1813 – 1662 (Print) E-ISSN: 2415 – 1726 (On Line)

17, 25, 27, 28). These results showed enough evidence that both IGRAs are not affected by the BCG vaccination which is reflected by high specificity of the two assays as they utilize specific antigens (ESAT 6 and CFP 10). The use of these two antigens provide a great improvement in the diagnosis of active TB as well as in the discrimination between MTB infection and former BCG vaccination. Therefore, the utilization of IGRAs could reduce the false diagnosis of MTB infection in particular BCG vaccinated and in non -MTB infected subjects.

*In conclusion*, IGRAs provide a supportive and complementary information as a part of the diagnostic work tools for tuberculosis. It is also important to note that a positive result confirms diagnosis of tuberculosis, but a negative IGRA does not rule it out. Of the two IGRAs QFT GIT is more convenient to be used.

testing to the diagnosis of latent tuberculosis infection in HIV infected patients: A comparison of Quanti FERON–Tb Gold in tube, T-SPOT TB and tuberculin skin test. BMC infectious Diseases. 12: 169.

**10.** Andersen, P., Munk ME, Pollock JM, and Doherty TM(2000). Specific immune-based diagnosis of tuberculosis. **Lancet 356 (9235):1099-1104.** 

**11.** Dunn OJ, Clark VA (2009). Basic statistics a primer for the biomedical sciences. **4th ed. John Wily and Sons, New Jersey. pp: 41-146.** 

12. Washigton WJ, Allen S, Janda W, Koneman E, Procop G and Schreckenberger P (2006). Koneman's Color atlas and textbook of diagnostic microbiology.  $6^{th}$  edition pp: 1461.

**13.** Pai M and Menzies D (2007). Interferon gamma release assay : what is their role in the diagnosis of active tuberculosis. **Clin Infect Dis**, **44**(1):74-77.

**14.** Detjen AK, Keil T, Roll S, Hauer B, Mauch H, Wahn U and Magdorf K (2007). Interferon gamma release assays improve the diagnosis of tuberculosis and nontuberculous mycobacterial disease in children in country with low incidence of tuberculosis. **Clin infect Dis. 45(3): 322-328.** 

**15.** Ruhwald M, Nakaoka H, Cuevas LE, Lawson L, Squire SB, Eugen-Olsen J and Ravn P (2008). Improving T-cells assays for the diagnosis of latent Tb infection :potential of a diagnosis test based on IP-10.**PLOS ONE. 3(8):e2858.** 

**16.** Abdel-Samea SA, Ismail YM, Fayed SM and Mohammad AA (2013). Comparative study between using QuantiFERON and tuberculin skin test in diagnosis of *Mycobacterium tuberculosis* infection. **Egyptian J of Chest Dis and Tuber. 62: 137-143.** 

**17.** Simsek H, Alpar SM, Aksu K, Aksu F, Ceyhan I, Cesur S, Gozalan A and Ertek M(2010).The comprehensive evaluation of latent tuberculosis infection in health care workers and patients with active tuberculosis using TST, ELISA and ELISPOT methods. **Turk J Med Sci. 40 (4): 585-591.** 

**18.** Sester M, Sotigiu G, Lange C, Giehl C, Migliori GB, Niehaus A, Ruhwald M, Wagner D, Zellweger JP, Huitric E, Sandgren A, Manissero D (2011). Interferon gamma release assays for the diagnosis of active tuberculosis: a systematic review and meta-analysis. **Eur Respir J. 37(1): 100-110.** 

**19.** Bianchi L, Galli L, Moriondo M, Veneruso G, Becciolini L, Azzari C, Chiappini E and de Martino M (2009). Interferon –gamma release assay improves the diagnosis of tuberculosis in children. **The pediatr Infect Dis J 28 (6): 510-514.** 

**20.** Franken WP, Timmermans JF, Prins C,Slootman HJ, Dreverman J, Bruins H, Dissel JT and Arend SM (2007). Comparison of mantoux and QuantiFERON Tb gold test for diagnosis of latent tuberculosis infection in Armey personnel. **Clin Vaccine Immunol 14 (4):477-480.** 

**21.** Palazzo R, Spensieri F, Massari M. (2008). Use of whole Samples in house bulk and single–cell antigen specific gamma interferon assay for surveillance of Mycobacterium tuberculosis infection. **Clin Vaccine Immunol 15:327-337.** 

**22.** Eddin LT and Monem F (2011). Utility of an interferon gamma release assay as a potential diagnostic aid for active pulmonary tuberculosis. **J Infect Dev Ctries. 6(1): 67-72.** 

**23.** Lee JY, Choi HJ,Park IN, Hong SB, Lim CM, Lee SD, Koh Y, Kim WS, Kim WD, Kim DS, Shim TS (2006). Comparison of two commercial interferon gamma assays for diagnosis of mycobacterial infection. **Eur Resp J 28: 24-30.** 

## ISSN: 1813 – 1662 (Print) E-ISSN: 2415 – 1726 (On Line)

24. Arend SM, Thijsen SF, Leyten EM, Bouwman JM, Franken WP, Koster BF, Cobclens FG, Houte AJ, Bossink WJ (2007). Comparison of two interferon gamma release assay and tuberculin skin test for tracing tuberculosis contacts. Am J Respir Crit Care Med. 175: 618-627.

**25.** Dominguez J, Manzano JR, De Souza-Galvao M, Latorre I. Milia C, Blanco S, Angeles-Jimenez M, Prat C, Lacoma A, Altet N, Ausin V (2008). Comparison of two commercially available gamma interferon blood tests for immunodiagnosis of tuberculosis. **Clin and Vaccine Immunol. 15(1): 168-171.** 

**26.** Horsburgh CR(2004). Priorities for the Treatment of Latent Tuberculosis Infection in the United States. **N Engl J Med.** 350:2060-2067.

27. Pai M, Gokhale K, Dogra S, Kalantri S, Mendiratta D, Mendiratta DK, Narang P, Daley CL, Granich RM, Mazurek GH, Reinglod L, Colford JM (2005). Mycobacterium tuberculosis infection in health care workers in rural India. JAMA. 8(22): 2746-2755.

**28.** De Souza- Galvao ML, Latorre I, Altet-Gomez N, Jimenez-Fuentes MA, Milia C, Solsona J, Seminario MA, Cantos A, Ruiz-Manzano J, Dominguez J (2014). Correlation between tuberculin skin test and IGRAs with risk factors for the spread of infection in close contacts with sputum smear positive in pulmonary tuberculosis. **BMC Infectious Diseases. 14:258.** 

المقارنة بين فحصي أطلاق الانترفيرون جاما في تشخيص التدرن الرئوي أسماء زكي شيتاوي آل – غرير ، زين العابدين عبدالعزيز عبدالله فرع الاحياء المجهرية ، كلية الطب ، جامعة الموصل ، العراق

#### الملخص

الأهداف: اختبار ومقارنة المقدرة التشخيصية لفحصي اطلاق الانترفيرون جاما (T SPOT TB and FERON Gold in Tube) في تشخيص التدرن الرئوي.

المواد وطريقة الدراسة: شملت الدراسة على 40 مريضا مصابا بالندرن الرئوي النشط الذين كانوا براجعون مراكز الرعاية الصحية النتفسية في مدينة الموصل للفترة من شهر آذار إلى شهر كانون الأول لعام 2013. تم جمع عينة قشع من كل مريض وزراعتها على الوسط الزرعي للفترة من شهر آذار إلى شهر كانون الأول لعام 2013. تم جمع عينة وشع من كل مريض وزراعتها على الوسط الزرعي الموصل للفترة من شهر آذار إلى شهر كانون الأول لعام 2013. تم جمع عينة وشع من كل مريض وزراعتها على 20 مريض الوسط الزرعي الموصل للفترة من شهر آذار إلى شهر كانون الأول لعام 2013. تم جمع عينة وشع من كل مريض وزراعتها على الوسط الزرعي الموصل للفترة من شهر آذار إلى شهر كانون الأول لعام 2013. تم جمع عينة وضعت في أنابيب اختبار تحوي على الهيبارين لغرض الحال وضعت في أنابيب اختبار تحوي على الهيبارين لغرض الحراء فحص 30 مريض وزلي الموسل الفرامي وراحية الموسل الموسل الموسل الموسل الفرعي وراحية الموسل الموسل الفرعي مع 2010 أول لي مريض أول لعام 2013. أول لعام 2013 أول لعام 2013. أول لعام 2013 أول لعام 2014. أول لعام 2014 أول لعام 2014 أول لعام 2014. أول لعام 2014 أول لعام 2014 أول لعام 2014 أول لعام 2014 أول لعام 2014. أول لعام 2014 أول لع

النتائج: من أصل 40 مريض ايجابي للصبغة المقاومة للحموضة 37(2.5%) كانت نتيجة الاستنبات ايجابية. وبالنسبة لدرجة خصوصية فحصي إطلاق الانترفيرون جاما وصلت إلى 100% وقدرت حساسية T-SPOT TB ب 91.89%. في حين بلغت حساسية QFT-GIT QFT- كانت القيمة التنبؤية الايجابية للفحصين 100%. اما القيمة التنبؤية السلبية لفحص T SPOT TB كانت50% بينما لفحص -QFT GIT كانت 37.5% كما أن درجة الاتفاق بين فحصي اطلاق الانترفيرون جاما كانت متوسطة(4.5%) هي حالة (82.5%) في حالة التدرن. كما الثبت أن نتيجة فحصي إطلاق الانترفيرون جاما لا تتأثر بلقاح BCG في مرضى التدرن.

الاستنتاجات: يمكن استخدام فحصى اطلاق الانترفيرون جاما كجزء من العملية التكميلية لتشخيص التدرن لكن من المهم ملاحظة ان النتيجة السالبة للفحصين لا يستبعدان الإصابة بمرض التدرن و أن من بين الفحصين, فحص QFT-GIT أكثر ملائمة للاستخدام.